



## Introduction—liquid biopsies in cancer studies

The term liquid biopsy is a fairly recent addition to the scientific literature (1,2) and was originally defined as “*A test made on a sample of blood to determine the presence of either circulating tumor cells (CTCs) or circulating DNA fragments (cfDNA) from a tumour circulating in the blood*”. Its importance lay in the minimally invasive approach needed for a blood sample for either CTC or cfDNA analysis and so could be repeated on the same patient at relatively frequent intervals and with minimum discomfort. It is also clear that this DNA may also be used in the monitoring of treatment (3,4) as well as prognosis.

Normally, a biopsy describes a small sample removed from a solid source [e.g., liver biopsy removed by fine needle aspiration (5)]. However, liquid samples can be removed routinely from a variety of sources from the patient in addition to blood [e.g., saliva, urine, cerebro-spinal fluid, peritoneal fluid, breast milk, tears, amniotic fluid, bronchial lavage aspirates and seminal fluid]. In many ways, therefore, the term liquid biopsy, should perhaps also be extended to include these other liquid sources employed for the identification of cfDNA and cfRNA (6) that form a basis for description by the term liquid biopsy so providing a relatively simple, minimally-invasive general approach. In particular, the use of blood for the detection of foetal cfDNA, first shown to be present by Lo (7), offers a minimally invasive approach to the diagnosis of foetal genetic defects and so a less dangerous approach than amniocentesis.

Liquid biopsy, in fact, has been in use for many years with perhaps the first example being that of Ashworth (8) when identifying CTCs in a human blood sample—though after death. The first example of its use for cfDNA and cfRNA in human blood sampled from a variety of patients is that of Mendel & Métais (9). The next big step towards the liquid biopsy concept came much later with the identification of cell-free tumour DNA (ctDNA) in human blood samples (10).

Although the initial definition limits the term to ctDNA in blood, it is also important to note that there is a range of components circulating in human blood including cfDNA/cfRNA from the breakdown of leucocytes, bacteria, viruses, mitochondria, cell and tissue necrosis, cell apoptosis, cellular release of exosomes, transposons, retrotransposons and viroplasm; parasite nucleic acids and leukocyte surface DNA. Of particular interest, in relation to the articles that follow, is the wide range of RNAs found in peripheral blood, namely: mRNA, tRNA, rRNA, snRNA, snoRNA, dsRNA, RNAi, siRNA, miRNA, piRNA, circRNA, ceRNA and lncRNA (11).

The term liquid biopsy should also be expanded to include studies on exosomes with their variety of RNA content. In addition, consideration should be given to actively released DNA (12-15) as opposed to that only released by apoptosis and necrosis.

The use of DNA liquid biopsies joins the similar studies started earlier on blood protein markers (16,17) and later on blood lipid markers (18). This has led to examining the combined use of potential DNA and protein markers for the detection of tumours and their treatment (19).

Examination of the technologies involved in liquid biopsies show their potential importance in the study of cancer and the various related aspects. The following articles in this special number may be considered in three parts, one dealing with CTCs, one with ctDNA and its associated histones whilst the third is based upon exosomes and the relevance of their RNA components.

CTCs can provide a useful guide to the genetic changes found in both the primary tumour and its metastases. The review on CTCs by Mai relates to their use in patient management whilst that by Riethdorf offers a resumé of various studies aiming to analyse miRNA expression in CTCs. Some of such miRNAs interfere with the post-translational activities of mRNA and may be involved with tumour development and progression. This could offer a way to a better understanding of the regulatory mechanisms involved in tumour cell dissemination and, possibly, the cellular origins of miRNAs.

Obviously, it is important to identify the presence of a tumour in a patient at an early stage and especially before it has had a chance to metastasise. Thus, Schmidt and Fleischhacker question the readiness of the use of liquid biopsy in early analysis. In particular, they consider the basic issues of handling the blood sample prior to analysis and the features that need to be controlled if liquid biopsy is to be successfully clinically applied across many different laboratories. Early detection is also a theme considered by Tanos and Thierry in their discussion of the clinical relevance of liquid biopsy for cancer screening and the usefulness of cfDNA, CTCs, cfRNAs as markers to form the basis of clinical tests.

The roles of ctDNA in early diagnosis, prognosis and the monitoring of treatment in cancer have been the subject of

many researches (20) since the identification of tumour DNA in blood (10). In particular, good progress has been made in the monitoring of treatment e.g., with the identification of patients capable of responding to (EGFR)-targeted monoclonal antibodies in colorectal cancer (21-23). Whilst well-studied, the identification of ctDNA markers for the early diagnosis of tumours has proved disappointingly slow. The inclusion of the study of liquid biopsy RNAs has added an additional approach that may have application in early diagnosis, treatment monitoring and prognosis (3).

Heitzer and colleagues examine the use of circulating tumour DNA that reflects the full spectrum of tumour-specific alterations. Many approaches to liquid biopsy involve targeted ctDNA fractions, although this may be considered to be an inadequate approach given the changing genetic landscape in tumours. Hence a review of untargeted approaches is offered to highlight the possible advantages—and limitations—of this approach over the targeted studies.

Size has become an important factor in cfDNA analysis for tumour markers with arguments existing in support of both the larger-sized versus the smaller fractions in early cancer detection (24,25). Grunt *et al.* describe below both methods and the various technical platforms by which cfDNA fractions are enriched and isolated for their identification of size profiles in plasma and serum.

Epigenetic changes have also been examined in cfDNA. Here, Rykova *et al.* consider investigations into cfDNA-based methylated markers including advances in analytical methods and their application to liquid biopsy cfDNA especially as applied to lung cancer diagnosis, treatment prediction and disease prognosis. Equally, Yörüker *et al.* discuss the post-translational modifications (PTMHs) to histone proteins that can affect gene expression and chromatin packaging. Enzyme systems involved in these processes can be affected in cancer cells so leading to changed global patterns of PTMHs in different cancers and hence their potential use as biomarkers.

Early studies indicated that unwanted RNAs could be destroyed by exosomes containing RNase (26) exosomes being found to contain many domains with RNAase activity. Thus, on release into the blood stream, they could affect the RNase levels. However, it is now clear that exosomes act importantly as vehicles for transferring a broad variety of RNAs between cells, so acting as an inter-cellular signalling system with important roles in both normal and pathological physiological activities. Their presence in a range of body fluids makes them useful clinically primarily through the range of RNAs, as well as DNA, in the signalling process, and hence as possible biomarkers in the tumour situation.

As with cfDNA, exosome preparation is of fundamental importance if the role(s) of the various RNAs in pathological conditions and their possible use as biomarkers can be replicated by all laboratories. Milica and de Marco discuss the inefficiency of the exosomal isolation and purification methods resulting, ultimately, to possible misleading results. They review the commonly-used approaches and consider their applicability for working with clinical material. Two forms of RNA found in exosomes are the non-coding microRNAs (miRNAs) and long non-coding RNAs (lncRNAs). The former is often deregulated in tumour cells leading to changes in the expression of tumour associated genes. The problem of standardized miRNA preparatory methods from plasma/serum of cancer patients is considered by Schwarzenbach. The other important form of exosomal RNA, lncRNA, is discussed by Dragomir *et al.* as it is an important regulator of physiological processes that is deregulated in cancer. These authors consider the action of exosomal lncRNAs in cancer as well as their possible interactive roles with other RNA species. The final contribution on exosomes by Meng *et al.* concerns not only exosomal roles in cancer and their possible use as biomarkers, but also exosomes as therapeutic target and drug delivery vehicles.

## Acknowledgments

*Funding:* None.

## Footnote

*Provenance and Peer Review:* This article was commissioned by the Guest Editor (Heidi Schwarzenbach) for the series “Technologies in Liquid Biopsies - Potential applications in Medicine” published in *Translational Cancer Research*. The article did not undergo external peer review.

*Conflicts of Interest:* The author has completed the ICMJE uniform disclosure form (available at <http://dx.doi.org/10.21037/>

tcr.2018.02.10). The series “Technologies in Liquid Biopsies - Potential applications in Medicine” was commissioned by the editorial office without any funding or sponsorship. The author has no other conflicts of interest to declare.

*Ethical Statement:* The author is accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

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## References

1. Crowley E, Di Nicolantonio F, Loupakis F, et al. Liquid biopsy: monitoring cancer-genetics in the blood. *Nat Rev Clin Oncol* 2013;10:472-84.
2. Perakis S, Speicher MR. Emerging concepts in liquid biopsies. *BMC Medicine* 2017;15:75.
3. Holdenrieder S. CNAPS in therapy monitoring. In: Gahan PB, editor. *Circulating Nucleic Acids in Early Diagnosis, Prognosis, and Treatment Monitoring. An introduction*. Springer Dordrecht 2015;325-67.
4. Berger AW, Schwerdel D, Welz H, et al. Treatment monitoring in metastatic colorectal cancer patients by quantification and KRAS genotyping of circulating cell-free DNA. *PLoS One* 2017;12:e0174308.
5. Diamantis A, Magiorkinis E, Koutselini H. Fine-needle aspiration (FNA) historical aspects. *Folia Histochem Cytobiol* 2009;47:191-7.
6. Hui L, Maron JL, Gahan PB. Other body fluids as non-invasive sources of cell-free DNA/RNA. In: Gahan PB, editor. *Circulating Nucleic Acids in Early Diagnosis, Prognosis, and Treatment Monitoring. An introduction*. Springer Dordrecht 2015;295-323.
7. Lo YM, Corbetta N, Chamberlain PF, et al. Presence of fetal DNA in maternal plasma and serum. *Lancet* 1997;350:485-7.
8. Ashworth TR. A case of cancer in which cells similar to those in tumours were seen in blood after death. *Aust Med J* 1869;14:146-9.
9. Mandel P, Métais P. Les acides nucléiques du plasma sanguine chez l'homme. *C R Seances Soc Biol Fil* 1948;142:241-3.
10. Stroun M, Anker P, Maurice P, et al. Neoplastic characteristics of the DNA found in the plasma of cancer patients. *Oncology* 1989;46:318-22.
11. Huang X, Yuan T, Tschannen M, et al. Characterization of human plasma-derived exosomal RNAs by deep sequencing. *BMC Genomics* 2013;14:319.
12. Stroun M, Anker P, Maurice P, et al. Circulating nucleic acids in higher organisms. *Int Rev Cytol* 1977;51:1-48.
13. Adams DH, Gahan PB. The DNA extruded by rat spleen cells in culture. *Int J Biochem* 1983;15:547-52.
14. Gahan PB, Stroun M. The virtosome—a novel cytosolic informative entity and intercellular messenger. *Cell Biochem Funct* 2010;28:529-38.
15. Bronkhorst AJ, Wentzel JF, Aucamp J, et al. Characterization of cell-free DNA released by cultured cancer cells. *Biochim Biophys Acta* 2016;1863:157-65.
16. Swisher EM, Wollan M, Mahtani SM, et al. Tumor-specific p53 sequences in blood and peritoneal fluid of women with epithelial ovarian cancer. *Am J Obstet Gynecol* 2005;193:662-7.
17. Thälén C, Lundström S, Seignez C, et al. Citrullinated histone H3 as a novel prognostic blood marker in patients with advanced cancer. *PLoS One* 2018;13:e0191231.
18. Viol-a-Magni MP, Cattaldi S, Marocco D. Bladder cancer markers and recent innovations. In: Ather MH, editor. *Bladder cancer - management of NMI and muscle-invasive cancer*. London, UK: INTECHOPEN Ltd, 2017.
19. Cohen JD, Javed AA, Thoburn C, et al. Combined circulating tumor DNA and protein biomarker-based liquid biopsy for the earlier detection of pancreatic cancers. *Proc Natl Acad Sci USA* 2017;114:10202-7.

20. Gahan PB, editor. *Circulating Nucleic Acids In Early Diagnosis, Prognosis, and Treatment Monitoring*. An introduction. Springer Dordrecht, 2015.
21. Heineman V, Douillard JY, Ducreux M, et al. Targeted therapy in metastatic colorectal cancer – An example of personalised medicine in action. *Cancer Treat Rev* 2013;39:592-601.
22. Santos C, Azuara D, Garcia-Carbonero R, et al. Optimization of RAS/BRAF mutational analysis confirms improvement in patient selection for clinical benefit to anti-EGFR treatment in metastatic colorectal cancer. *Mol Cancer Ther* 2017;16:1999-2007.
23. Thierry AR, El Messaoudi S, Mollevi C, et al. Clinical utility of circulating DNA analysis for rapid detection of actionable mutations to select metastatic colorectal patients for anti-EGFR treatment. *Ann Oncol* 2017;28:2149-59.
24. Moulière F, Robert B, Peyrotte EA, et al. High fragmentation characterizes tumour-derived circulating DNA. *PLoS One* 2011;6:e23418.
25. Moulière F, Rosenfeld N. Circulating tumour-derived DNA is shorter than somatic DNA in plasma. *Proc Nat Acad Sci USA* 2015;112:3178-9.
26. Alberts B, Johnson A, Lewis J, et al. *Molecular biology of the cell*. 4th edition. New York: Garland Scientific Publishing, 2002.



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Submitted: 30 January 2018; Accepted for publication: 6 February 2018

doi: 10.21037/tcr.2018.02.10

**View this article at:** <http://dx.doi.org/10.21037/tcr.2018.02.10>

**Cite this article as:** Gahan PB. Introduction—liquid biopsies in cancer studies. *Transl Cancer Res* 2018;7(Suppl 2):S101-S104. doi: 10.21037/tcr.2018.02.10