



The efficacy of Raman spectroscopy in the diagnosis of esophageal cancer: a systematic review and meta-analysis

Jianqi Hao^{1,2}, Cong Chen^{1,2}, Hongyu Jin², Nan Chen^{1,2}, Jian Zhou^{1,2}, Yuzhou Zhu², Kayi Chung², Qiang Pu^{1,3}

¹Department of Thoracic Surgery, West China Hospital, Sichuan University, Chengdu, China; ²West China School of Medicine, Sichuan University, Chengdu, China; ³Western China Collaborative Innovation Center for Early Diagnosis and Multidisciplinary Therapy of Lung Cancer, Sichuan University, Chengdu, China

Contributions: (I) Conception and design: Q Pu, J Hao; (II) Administrative support: Q Pu; (III) Provision of study materials or patients: H Jin, K Chung; (IV) Collection and assembly of data: C Chen, Y Zhu; (V) Data analysis and interpretation: J Hao, J Zhou, N Chen; (VI) Manuscript writing: All authors; (VII) Final approval of manuscript: All authors.

Correspondence to: Dr. Qiang Pu. Department of Thoracic Surgery, West China Hospital, Sichuan University, Chengdu, China. Email: puqiang100@163.com.

Background: Esophageal cancer is characterized by its high occurrence rate and difficulty to treat successfully, therefore early diagnosis is extremely important. Our article is aimed to comprehensively analyze the relative effectiveness of Raman spectroscopy (RS) in the diagnosis of suspected esophageal cancer.

Methods: We performed a systemic search in PubMed, EMBASE, CNKI and Web of science from 2007 to 2020. We used the diagnostic data of true positive (TP), false positive (FP), true negative (TN) and false negative (FN) to estimate the pooled sensitivity, specificity, positive and negative likelihood ratios (LR), and diagnostic odds ratio (DOR), with 95% confidence intervals (CI), based on random effects models. The SROC curve analysis and AUC were performed to estimate the overall performance of Raman spectroscopy. QUADAS-2 guidelines were used to evaluate the quality of each study.

Results: Nine articles were included according to our inclusion and exclusion criteria. General pooled diagnostic sensitivity and specificity of RS to esophageal cancer were 0.91 (95% CI, 0.89–0.93) and 0.92 (95% CI, 0.91–0.94). The pooled PLR and NLR were 18.98 (95% CI, 6.61–54.49) and 0.09 (95% CI, 0.05–0.16), respectively. The DOR was 217.21 (95% CI, 68.32–690.53) indicating high accuracy. And the AUC of SROC curve was 0.9779.

Conclusions: We found a promisingly high sensitivity and specificity of RS in the diagnosis of suspected esophageal mass.

Keywords: Raman spectroscopy; diagnostic efficacy; esophageal cancer

Submitted Feb 05, 2020. Accepted for publication Jul 08, 2020.

doi: 10.21037/tcr-20-854

View this article at: <http://dx.doi.org/10.21037/tcr-20-854>

Introduction

Esophageal cancer is a malignant lesion formed by exceed proliferation of esophageal squamous epithelium or glandular epithelium. Esophageal cancer is the eighth most common malignant tumors in the world (1,2), which is characterized by its high occurrence rate and difficulty to

treat successfully following the thoracic surgery. Especially in China, the incidence of esophageal cancer ranks fifth with an average annual death toll of about 150,000 (3,4). The treatment of early esophageal cancer is comparatively simple, and the postoperative survival rate is also high. The 5-year survival rate of early esophageal squamous cell carcinoma is 62.9–92.6%, and the advanced stage is only

about 30% (5,6). However, early esophageal cancer often has no symptoms. When symptoms such as swallowing obstruction appearing, it is always advanced. It suggests that early diagnosis of esophageal cancer from routine physical examination will undoubtedly have important significance for the treatment of patients with esophageal cancer (7). In order to enhance early diagnosis and treatment, more frequent use is imaging techniques, especially thoracic computed tomography (CT). For patients with suspected esophageal masses, who expect long-term surveillance or non-surgical interventions, an esophageal mass biopsy (EMB) is usually recommended to observe pathological features to guide subsequent treatment options (8). However, a number of factors may have an impact on EMB diagnostic accuracy, including the cut-off number of specimens and the interpretive degrees of pathologists (9). Accordingly, the diagnostic accuracy of EMB varies from 79% to 100%, important clinical diseases easy to be incorrectly identified including fibrosis and necrosis (10). Therefore, a more trustworthy diagnostic method is urgently necessary.

Recently, it has reported that Raman spectroscopy (RS) has been applied clinically to determine the benign and malignant essence of tumor in surgeries for its ability to optically characterize the internal compositional properties (11,12). What's more, RS examination can be carried out *ex* or *in vivo* and it also can be a real-time, label-free and nondestructive (13). Theoretically, RS detects a variation of wave-length or Raman shift resulted from the inelastic light scattering from certain molecules (14). Different molecules have distinct combinations of Raman shifts which can produce unique spectral signatures (15). Therefore, Raman spectra are markedly related to the internal compositional features of tissues of different properties. Importantly, the availability to be performed *in vivo*, label-free, real-time and non-destructively perfectly addresses the deficiencies of traditional EMB. In the past decade, many studies concerned on the accuracy, sensitivity, and specificity of RS in determining the quality of unknown esophageal mass varied widely from one another, and some studies failed to recruit a sufficient number of patient samples, which could lead to potential bias and inaccuracy (16). Therefore, in order to comprehensively analyze the accurate diagnostic efficiency of RS in determining the benign and malignant features of esophageal tumors, mainly on the parameters of sensitivity, specificity and accuracy, we conducted meta-analysis and systematic evaluation to determine the value of clinical RS. We present the article in accordance with

the PRISMA reporting checklist (available at <http://dx.doi.org/10.21037/tcr-20-854>).

Methods

Literature research

Two independent reviewers conducted a systemic search through PubMed, EMBASE, CNKI and Web of science. The combinations of the relevant medical subject heading (MeSH) terms, key words, and word variants for “esophagus”, “esophageal”, “Raman spectra” were performed for the primary search. All available studies were published up to January 2020 with no other special limitation, if any discordance happened, we resolved it by consensus. In addition to searching online and in order to identify potentially eligible articles, we also hand-searched the bibliographies of review articles.

Selection criteria and exclusion criteria

As a meta-analysis, articles like single sample experiment, comments, case report, letters review articles and editorials were eliminated from the study. Finally, remaining studies were carefully selected when meeting the significant criteria as follows: (I) without animal tissues in the experiments; (II) reported the use of RS in esophageal cancer; (III) recruiting time from January 2007; (IV) used histopathology to confirm the diagnosis; (V) reported the true positive (TP), false positive (FP), true negative (TN) and false negative (FN), based on which the sensitivity and specificity values can be calculated.

Exclusion criteria: (I) studies that involved nonhuman subjects, (II) studies with no relevant data on diagnostic performance; (III) studies like case reports and case series were excluded. If a potential discrepancy was detected, a blinded third reviewer was assigned to adjudicate the conflict.

Data extraction

The parameters were extracted by two independent experienced investigators with a standard extraction table, if we had any confliction, we would launch a discussion to reach a consensus. The listed information of the essays was extracted basic information like title, author, nationality and enrolled year. Then the primary parameters, which indicated the diagnostic value, including TP, FP, TN and

FN were extracted from all the included studies.

Statistical analysis

The diagnostic value of RS for esophageal cancer was evaluated by using the primary data of TP, FP, TN and FN. Then pooled sensitivity, specificity, positive and negative likelihood ratios (LR), and diagnostic odds ratio (DOR) were integrated by random effects models with their P values and 95% CIs. Sensitivity and specificity were shown in forest plots using Meta-Disc version 1.4 statistical software.

Meanwhile, combination of sensitivity and specificity were assessed by summary receiver operator characteristics (SROC). The overall diagnostic accuracy of RS was estimated by the area under the curve (AUC) of SROC. An AUC value of greater than 0.9 defined a diagnostic tool as excellent, while between 0.8 and 0.9 as good, between 0.7 and 0.8 as fair and less than 0.7 as poor. The SROC curves were also made through Meta-Disc version 1.4. At the same time, according to the detection method *in vivo* and *in vitro*, we conducted a subgroup analysis.

Quality assessments and publication bias

We evaluated the quality of included studies through the Quality Assessment of Diagnostic Accuracy Studies (QUADAS-2) guidelines by Review Manager 5.3. We evaluated publication bias through Deeks Funnel Plot Asymmetry Test (consider the existence of publication bias when $P < 0.05$) by Stata 14.2 (StataCorp, USA).

Results

Search results

The initial process of study screening was shown in *Figure 1*. A total of 69 potential correlative articles were searched in PubMed and other databases in January 10th 2020, finally 9 articles (17-24) were eligible on the basis of our criterion.

Characteristics of the included studies

The general characteristics of the 9 eligible articles were summarized in *Table 1*. After searching the authenticated databases, article screening and quality assessment process, 9 articles with high quality and reliability, and comprehensible design with full texts and accessible data were considered

for this systematic review. All included researches were published in English except two which was published in Chinese. The total number of patients incorporated was 695 with one study didn't report its number of patients, and the total number of spectra incorporated was 3,834. And specimens were collected from patients from January 2007 to January 2020. Diagnostic algorithm includes leave-one patient-out, cross validation (LOPCV) in one article, principal components analysis (PCA) in one article, linear discriminate analysis (LDA) in three articles and partial least squares-discriminant analysis (PLS-DA) in two articles. In terms of the nationalities, 4 studies were from China ($n=4$), 3 the studies were from the same team of Singapore ($n=3$) and others were performed in Japan ($n=1$) and England ($n=1$). Of all the studies, 3 studies detected the esophageal mass by *in vivo* tissues, and 4 studies detected the esophageal mass by *ex vivo* tissues, whereas the other two studies were measured from urine and hemoglobin. Histopathology was used as the golden standard to confirm the diagnosis.

Pooled results

Among the nine studies, three studies used RS to screen suspected Barrett's esophagus. However, only one of the three studies reported the date of TN, FN, TP, FP. So, we cannot conduct a subgroup analysis about the diagnosis of Barrett's esophagus. Additionally, all the nine studies used RS to tell apart the benign and malignant feature of a particular tissue during an operation. And since some studies were conducted *ex vivo* and other studies were performed *in vitro*. Thus, we did the subgroup evaluation, which was divided into *ex vivo* and *in vitro* groups.

General pooled data

The sensitivity of the seven included articles which used RS to screen esophageal cancer with a particular tissue during an operation, ranged from 0.81 (95% CI, 0.66–0.91) in a study with 91 samples to 0.97 (95% CI, 0.94–0.99) which recruited 48 patients (1,172 spectra). The pooled sensitivity was 0.91 (95% CI, 0.89–0.93), which indicated a relatively low incidence rate of missed diagnosis. Particularly, among the seven included studies, except for one with sensitivity of 81%, the other six studies all maintained a sensitivity more than 85%. The forest plot of pooled sensitivity of all the seven studies was shown in *Figure 2A*.

The specificity of the seven studies ranged from 0.85 (95% CI, 0.82–0.87) in a study with 373 patients (200



PRISMA 2009 Flow Diagram

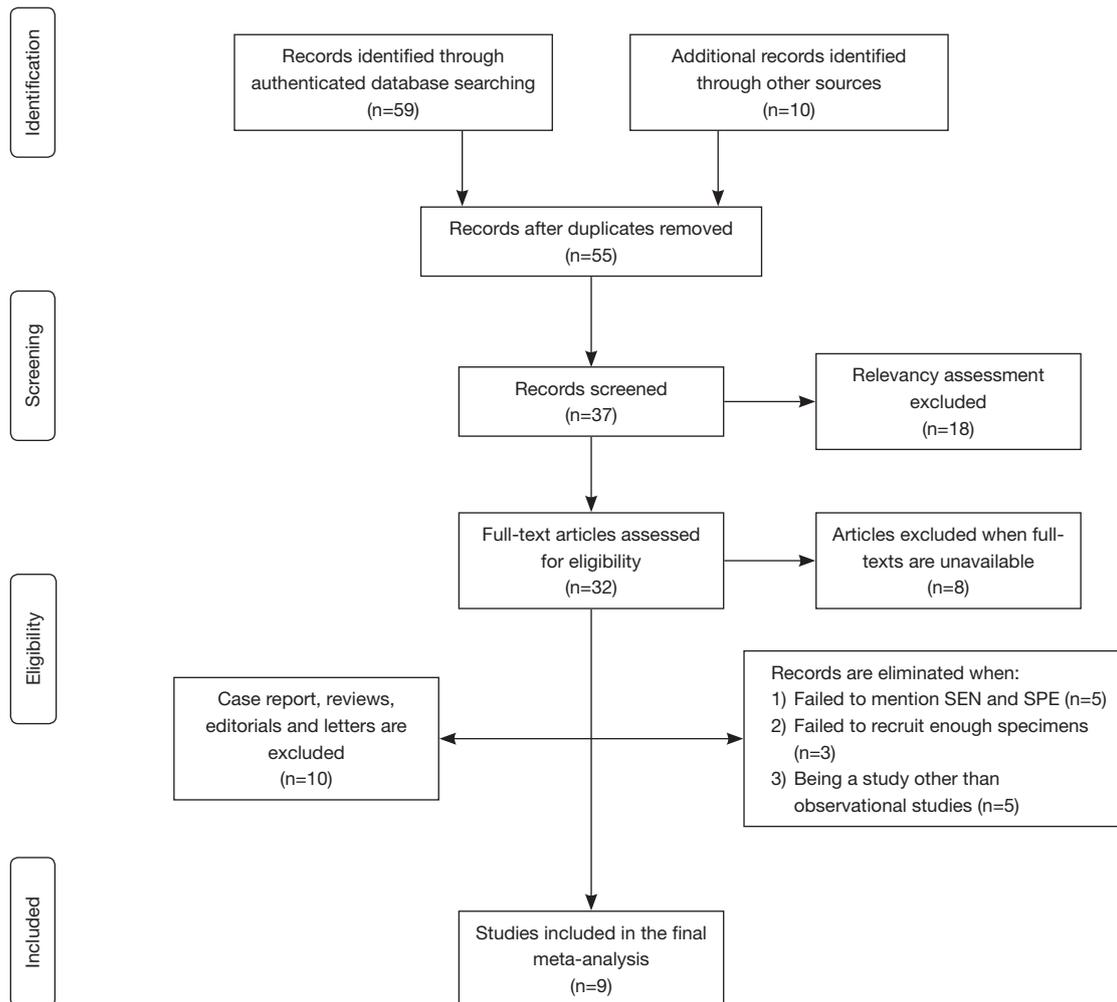


Figure 1 PRISMA 2009 flow diagram.

spectra) to 1.00 (95% CI, 0.94–1.00) by studies with 64 patients (128 spectra). The general pooled specificity was 0.92 (95% CI, 0.91–0.94), which was also a satisfactory parameter indicating a comparatively low rate of incorrect diagnosis. The forest plot of pooled specificity of all the seven studies was shown in *Figure 2B*.

The pooled PLR and NLR were 18.98 (95% CI, 6.61–54.49) and 0.09 (95% CI, 0.05–0.16), respectively. The DOR was 217.21 (95% CI, 68.32–690.53) indicating high

accuracy. The overall diagnostic accuracy was evaluated through the SROC curve analysis. And the AUC of the SROC curve was 0.9779. The plots were shown in *Figure 2C*.

Subgroup analysis

Ex vivo group

Of all the included studies, five studied conducted an

Table 1 Baseline characteristics of included studies

Reference	Country	N1	N2	N3	Age	Diagnostic algorithm	TP	FP	FN	TN	Sensitivity	Specificity	Vivo or vitro	Spectra	Accuracy
Almond, 2013	UK	62	673	798	NA	LDA	172	10	27	241	86%	88%	Vitro	830 nm	U
Bergholt, 2011	Singapore	107	207	1189	66	LDA	30	14	3	216	91%	93.9%	Vivo	785 nm	94.7%
Bergholt, 2014	Singapore	373	NA	200	NA	U	67	110	10	610	87%	85%	Vivo	785 nm	90%
Feng, 2016	China	169	NA	149	NA	PLS-DA	55	0	0	32	100%	100%	Vitro	785 nm	100%
Ishigaki, 2015	Japan	NA	91	NA	NA	LDA	34	3	8	47	81%	94%	Vitro	785 nm	U
Wang, 2015	China	48	U	1172	U	PLS-DA/LOPCV	196	19	6	717	97%	97.4%	Vivo	785 nm	U
Bergholt, 2011	Singapore	27	75	75	65	U	32	2	1	40	97%	95.2%	Vivo	785 nm	96%
Jiang, 2007	China	64	128	128	56	U	61	0	3	64	95.31%	100%	Vitro	785 nm	U
Zhou, 2013	China	41	41	123	U	PCA	19	1	2	19	90.48%	95%	Vitro	785 nm	U

U, unknown; N1, number of patients; N2, number of samples; N3, number of spectra; PCA, principal component analysis; LDA, linear discriminate analysis; PLS-DA, partial least squares-discriminant analysis; LOPCV, leave-one patient-out, cross validation.

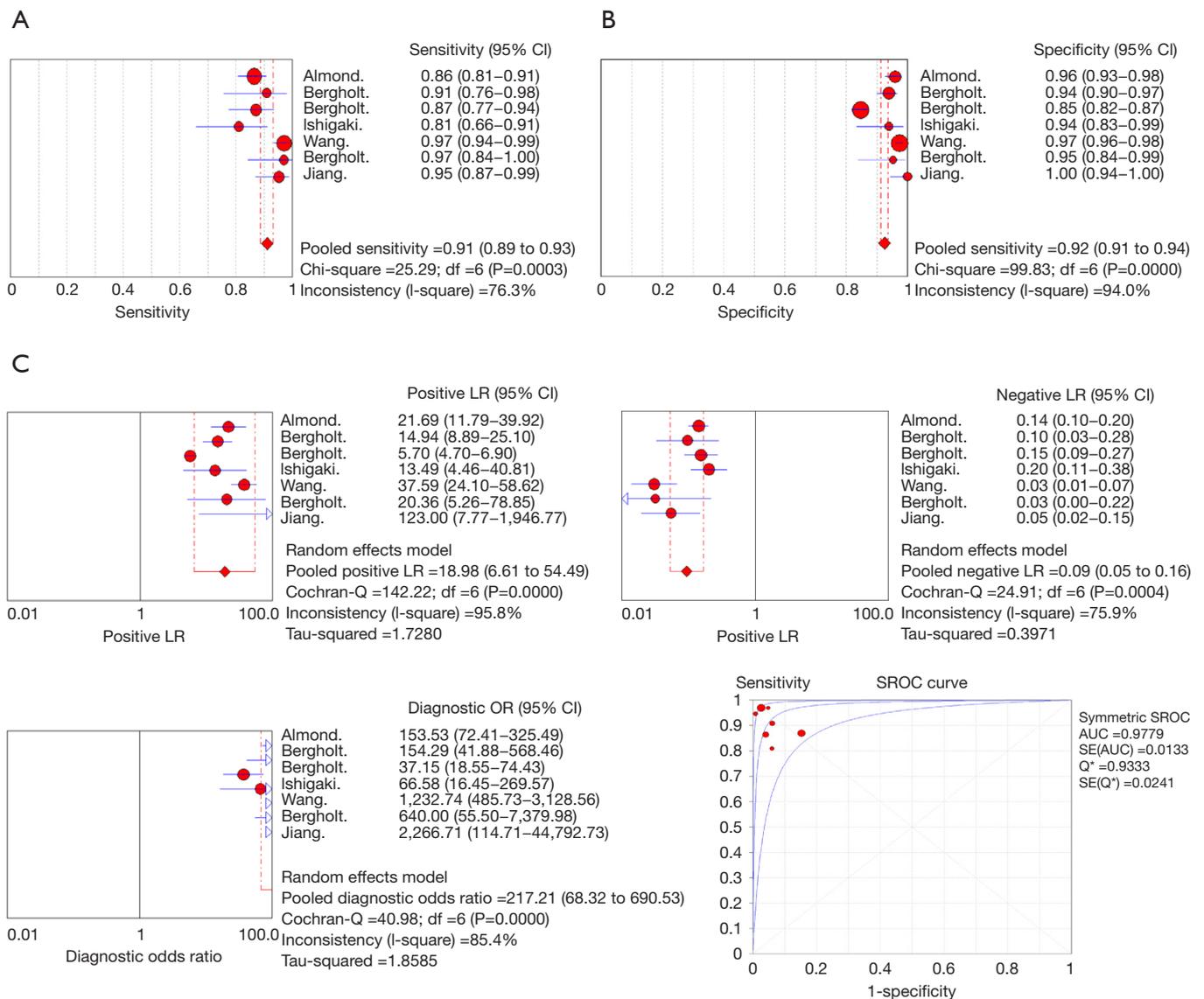


Figure 2 The pooled diagnostic efficacy of Raman spectroscopy in Esophageal cancer. (A) The forest plot of pooled sensitivity and their 95% confidence intervals (CIs) of Raman spectroscopy to diagnose esophageal cancer of all the nine studies. (B) The forest plot of pooled specificity and their 95% confidence intervals (CIs) of Raman spectroscopy to diagnose esophageal cancer of all the nine studies. (C) The pooled PLR, NLR, DOR and SROC curve of Raman spectroscopy in diagnosis of esophageal cancer. PLR, positive likelihood ratios; NLR, negative likelihood ratios; DOR, diagnostic odds ratios; SROC, summary receiver operator characteristics.

examination of esophageal specimens *in vivo*. The sensitivity of RS for esophageal specimens ranged from 0.81 (95% CI, 0.66–0.91) to 1.00 (0.94–1.00) and the pooled sensitivity was 0.94 (95% CI, 0.91–0.96). The specificity of RS for esophageal specimens ranged from 0.94 (95% CI, 0.83–0.99) to 1.00 (95% CI, 0.89–1.00) and the pooled specificity was 0.96 (95% CI, 0.94–0.98). The overall PLR and NLR were 20.16 (95% CI, 12.56–32.38) and 0.12 (95% CI, 0.06–0.22),

respectively. The DOR was 186.77 (95% CI, 69.63–500.93). The SROC curve analysis was also used to evaluate the overall diagnostic accuracy. And the AUC was 0.9912. All of the plots of *ex vivo* group were shown in *Figure 3*.

In vivo group

Of all the included studies, four studied conducted an examination of esophageal specimens *in vivo*. The pooled

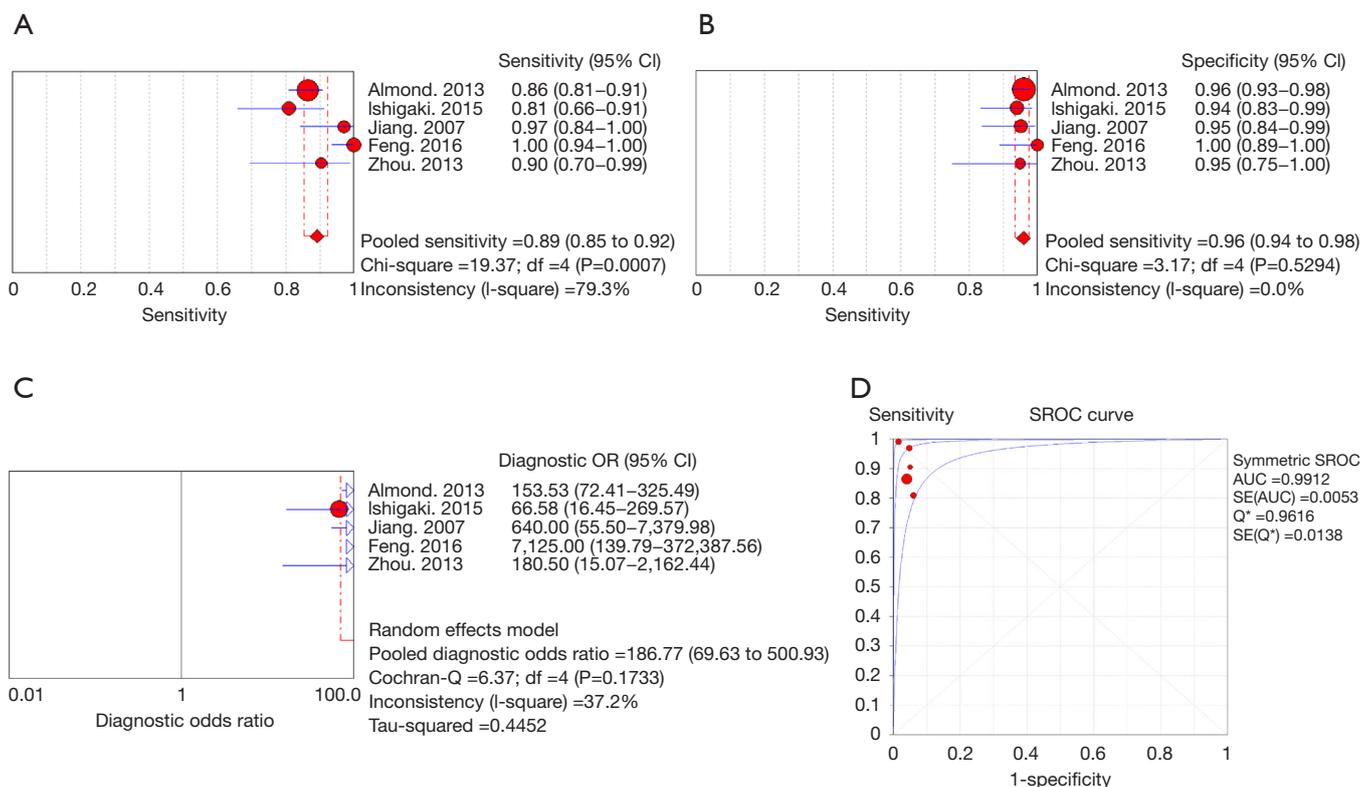


Figure 3 The plots of sensitivity, specificity, DOR and SROC curve of Raman spectroscopy in diagnosis of esophageal cancer *ex vivo*. DOR, diagnostic odds ratios; SROC, summary receiver operator characteristics.

sensitivity was 0.94 (95% CI, 0.91–0.96) ranged between 0.87 (95% CI, 0.77–0.94) and 0.97 (95% CI, 0.94–0.99). The pooled specificity was 0.92 (95% CI, 0.90–0.93) ranged between 0.85 (95% CI, 0.82–0.87) and 0.97 (95% CI, 0.96–0.98). The overall PLR and NLR were 15.70 (95% CI, 3.90–63.23) and 0.07 (95% CI, 0.03–0.18), respectively. The DOR was 214.67 (95% CI, 33.71–1,732.40). The SROC curve analysis was also used to evaluate the general diagnostic accuracy. And the AUC was 0.9812. All of the plots of *in vivo* group were shown in *Figure 4*.

Quality assessment and publication bias

Standard quality evaluation of the 9 included studies were performed based on QUADAS-2 by Review Manager 5.3. Also, publication bias was evaluated through Deeks Funnel Plot Asymmetry Test (consider the existence of publication bias when $P < 0.05$) which was conducted by Stata 14.2 (StataCorp, USA). The QUADAS-2 graphical display of the evaluation of the risk of bias was shown in *Figure 5*. The Deeks' funnel plot asymmetry test was used to assess

publication bias and it demonstrated that no significant publication bias was found ($P = 0.52$), which was shown in *Figure 6*.

Discussion

This meta-analysis was performed on the basis of the standard protocol for a systematic review with nine articles were taken into account. Two independent reviewers were assigned in study screening, data extraction and quality assessment process. The SROC curve analysis was simultaneously applied. We expect to assess the value of Raman spectroscopy in diagnosing esophageal cancer and its value in clinical application.

This meta-analysis gave an evidence that RS had a good diagnostic accuracy in esophageal cancer, with the general pooled diagnostic sensitivity and specificity being 0.91 (95% CI, 0.89–0.93) and 0.92 (95% CI, 0.91–0.94). According to the general pooled data, we found that the overall sensitivity and specificity were over 90%, a high efficacy of early diagnosis of esophageal cancer was reconfirmed

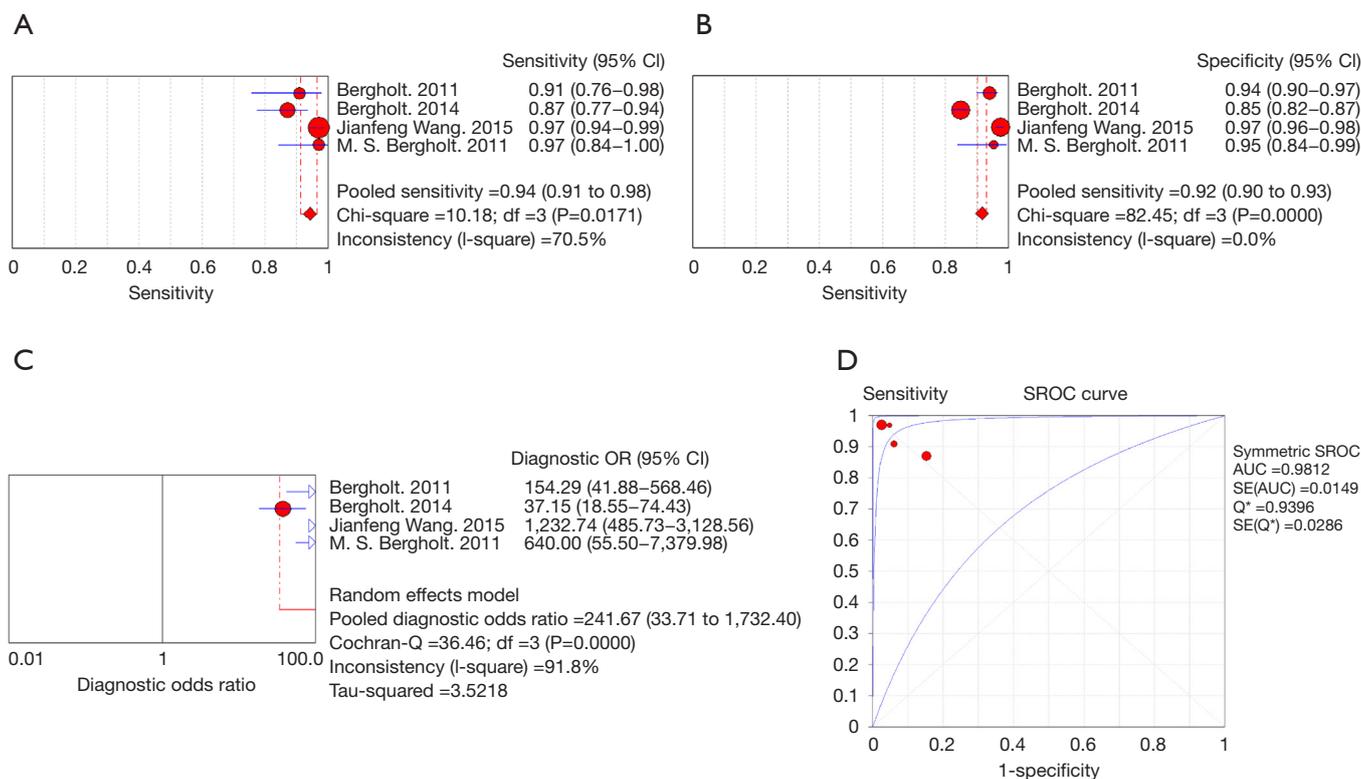


Figure 4 The plots of sensitivity, specificity, DOR and SROC curve of Raman Spectroscopy in diagnosis of esophageal cancer *in vivo*. DOR, diagnostic odds ratios; SROC, summary receiver operator characteristics.

which suggested that RS had a low missed diagnosis rate in distinguishing between benign and malignant aspects of esophageal cancer. Moreover, we also found that the general pooled DOR was (95% CI, 68.32–690.53) by random effect model, indicating a high accuracy. In SROC analysis, AUC was 0.9779. Generally, the value of AUC is lying between 0.5 and 1.0, and the larger AUC indicates better performance. In the study, the AUC value was very close to 1, which suggested an excellent diagnostic efficiency. In addition, four of the nine included studies reported the accuracy of the RS in diagnosis of esophageal cancer range from 90% to 100%, also confirmed our conclusion.

In the subgroup analysis, the overall sensitivity and specificity of the diagnosis exceeded 90%, whether Raman spectroscopy analyze esophageal cancer tissues *in vivo* or *in vitro* both showed high diagnostic accuracy. Compared to intraoperative biopsy, the diagnosis of Raman spectroscopy might have the same diagnostic performance, and with an irreplaceable function that frozen biopsy cannot reach. Furthermore, in the subgroup analysis, we found that two studies did not use intraoperative masses to verify

the benign and malignant esophageal cancer. They used hemoglobin and urinary modified nucleotides to diagnose esophageal cancer *in vitro*, so we didn't put them into general analysis.

In the study of Zhou (24), by detecting the hemoglobin solution of 21 patients with esophageal cancer and 20 healthy people, they found that the RS of hemoglobin in patients with esophageal cancer and healthy people were significantly different. According to the analysis of characteristic peaks, they reported that the contents of tyrosine, phenylalanine and the number of vibration of pyrrole ring in hemoglobin of patients with esophageal cancer were slightly lower than those of healthy people. Meanwhile, compared with healthy people, esophageal cancer patients had the increased iron ions in low spin state in hemoglobin, while the iron ions in high spin state decreased, which indicated that the iron ions in high spin state in hemoglobin of esophageal cancer patients were transferred to low spin state, which was consistent with the phenomenon that the blood samples of cancer patients were more easily hemolyzed. Hence, they believe that Raman

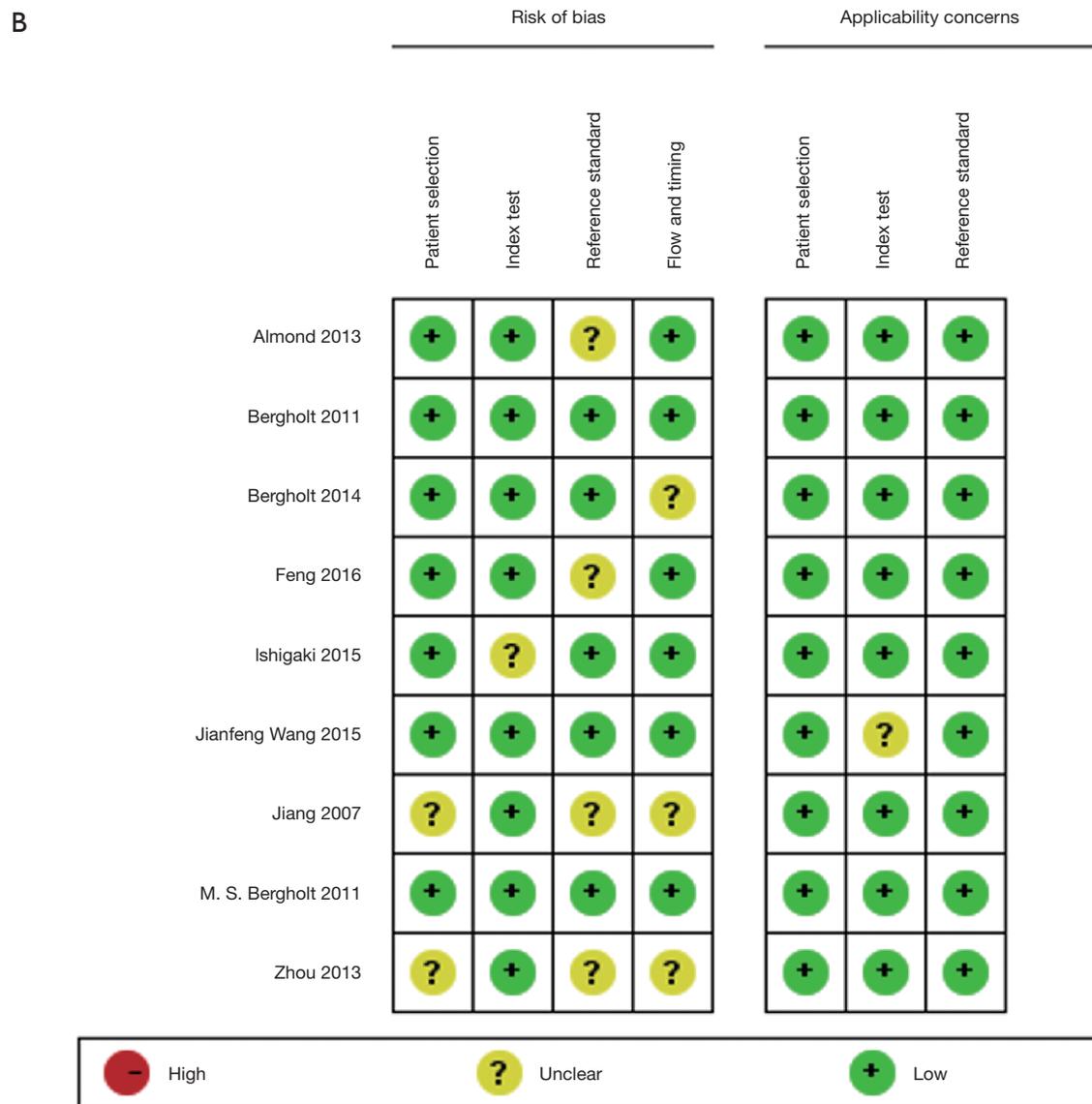
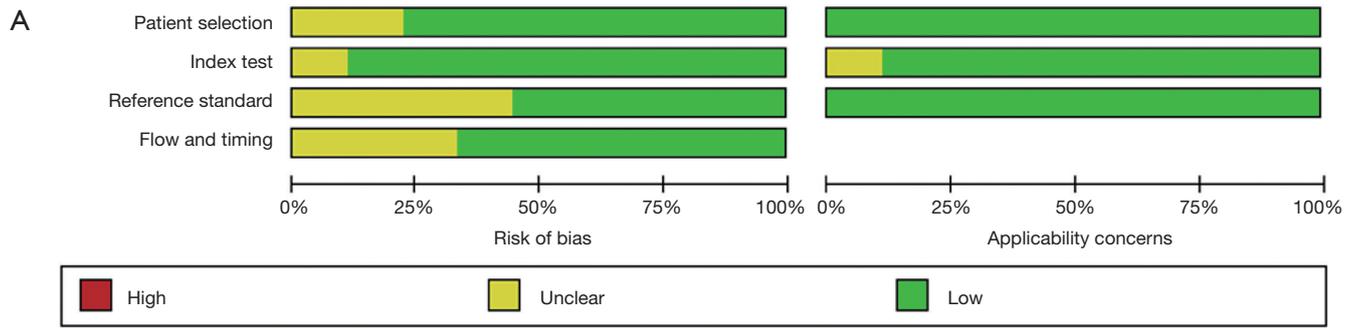


Figure 5 The graphical display of the evaluation of the risk of bias and concerns regarding applicability of the selected studies. (A) Risk of bias and applicability concerns evaluation of included studies in pool. (B) Risk of bias and applicability concerns evaluation of included studies individually.

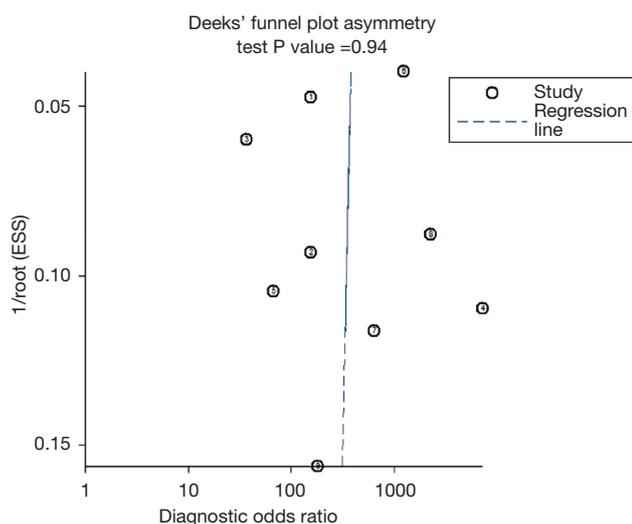


Figure 6 The Deeks' funnel plot asymmetry test.

spectroscopic analysis of hemoglobin might become a new tool for early diagnosis of esophageal cancer. Their research provided a new idea for the early diagnosis of esophageal cancer, but due to the lack of relevant research at present, more clinical studies were needed to verify this inference.

In the study of Feng (20), they used RS to diagnose the benign and malignant of esophageal cancer by urine. As one of the most noteworthy bio-fluids, human urine contains many metabolites which provide abundant diagnostic information about the human health status (25). In the study of Feng, they found that the average RS peaks of esophageal cancer group at 765 and 1,184 are more dense than normal group, while that of esophageal cancer group RS peaks at 725, 863, and 1,475 are lower. This information indicates the diagnostic potential of RS for differentiation of esophageal cancer.

Today, liquid biopsy technology has become a popular research in the diagnosis of various diseases (26). Raman spectroscopy has a unique role in liquid diagnostics which demonstrated that using RS in liquid diagnostics for diagnosing cancer might be a convenient potential method. It may promote the application of RS in cancer diagnosis.

A large number of studies have shown that RS does have high diagnostic specificity and sensitivity in the diagnosis of esophageal cancer (27). However, very few devices are actually used in clinical practice. We think it may be due to the fact that RS equipment is relatively expensive, and the sample preparation requirements are high (11,28), making it a low application rate. If the Raman spectroscopy device can

be improved in the future, it will be popularized. It will help early diagnosis of esophageal cancer patients, and it may also improve the survival of patients with esophageal cancer.

In our meta-analysis, we reconfirmed that RS had an excellent diagnostic efficiency in esophageal cancer. Simultaneously, we acknowledged several limitations in this study. Firstly, RS had not been widely admitted as a normal clinical diagnostic tool, therefore inadequate number of clinical researches were published, which absolutely lowered the number of articles we could include. Secondly, since most of the included researches were conducted in Asia, it might cause selection bias. Thirdly, due to limitations of current research, our study did not involve tumor subtypes. Further comprehensive study is needed so that it can target the subtypes in order to provide more precise clues for clinical practice.

Conclusions

Through our meta-analysis, we found a promisingly high sensitivity and specificity of RS in the diagnosis of suspected esophageal mass.

Acknowledgments

Funding: Supported by the Major Scientific and Technological Projects of the New Generation of Artificial Intelligence in Sichuan Province in 2018 under Grant 2018GZDZX0035.

Footnote

Peer Review File: Available at <http://dx.doi.org/10.21037/tcr-20-854>

Reporting Checklist: The authors have completed the PRISMA reporting checklist. Available at <http://dx.doi.org/10.21037/tcr-20-854>

Conflicts of Interest: All authors have completed the ICMJE uniform disclosure form (available at <http://dx.doi.org/10.21037/tcr-20-854>). The authors have no conflicts of interest to declare.

Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are

appropriately investigated and resolved.

Open Access Statement: This is an Open Access article distributed in accordance with the Creative Commons Attribution-NonCommercial-NoDerivs 4.0 International License (CC BY-NC-ND 4.0), which permits the non-commercial replication and distribution of the article with the strict proviso that no changes or edits are made and the original work is properly cited (including links to both the formal publication through the relevant DOI and the license). See: <https://creativecommons.org/licenses/by-nc-nd/4.0/>.

References

1. Siegel R, Ma J, Zou Z, et al. Cancer statistics, 2014. *CA Cancer J Clin* 2014;64:9-29.
2. Bray F, Jemal A, Grey N, et al. Global cancer transitions according to the Human Development Index (2008-2030): a population-based study. *Lancet Oncol* 2012;13:790-801.
3. The L. GLOBOCAN 2018: counting the toll of cancer. *Lancet* 2018;392:985.
4. Bray F, Ferlay J, Soerjomataram I, et al. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin* 2018;68:394-424.
5. Zeng H, Zheng R, Zhang S, et al. Esophageal cancer statistics in China, 2011: Estimates based on 177 cancer registries. *Thorac Cancer* 2016;7:232-7.
6. Wang GQ, Jiao GG, Chang FB, et al. Long-term results of operation for 420 patients with early squamous cell esophageal carcinoma discovered by screening. *Ann Thorac Surg* 2004;77:1740-4.
7. Smyth EC, Lagergren J, Fitzgerald RC, et al. Oesophageal cancer. *Nat Rev Dis Primers* 2017;3:17048.
8. Kato H, Nakajima M. Treatments for esophageal cancer: a review. *Gen Thorac Cardiovasc Surg* 2013;61:330-5.
9. Chen D, Goldblum JR, Landau M, et al. Semiquantitative histologic evaluation improves diagnosis of esophageal carcinoma cuniculatum on biopsy. *Mod Pathol* 2013;26:806-15.
10. Storch I, Shah M, Thurer R, et al. Endoscopic ultrasound-guided fine-needle aspiration and Trucut biopsy in thoracic lesions: when tissue is the issue. *Surg Endosc* 2008;22:86-90.
11. Eberhardt K, Stiebing C, Matthaus C, et al. Advantages and limitations of Raman spectroscopy for molecular diagnostics: an update. *Expert Rev Mol Diagn* 2015;15:773-87.
12. Butler HJ, Ashton L, Bird B, et al. Using Raman spectroscopy to characterize biological materials. *Nat Protoc* 2016;11:664-87.
13. Garai E, Sensarn S, Zavaleta CL, et al. A real-time clinical endoscopic system for intraluminal, multiplexed imaging of surface-enhanced Raman scattering nanoparticles. *PLoS One* 2015;10:e0123185.
14. Esmonde-White KA, Cuellar M, Uerpmann C, et al. Raman spectroscopy as a process analytical technology for pharmaceutical manufacturing and bioprocessing. *Anal Bioanal Chem* 2017;409:637-49.
15. Lu F, Huang T, Han L, et al. Tip-Enhanced Raman Spectroscopy with High-Order Fiber Vector Beam Excitation. *Sensors (Basel)* 2018;18:3841.
16. Li X, Lin J, Jin H, et al. Spectral analysis for diagnosis of esophagus dysplasia using fluorescence Raman spectroscopy. *Conf Proc IEEE Eng Med Biol Soc* 2004;2006:141-4.
17. Almond LM, Hutchings J, Lloyd G, et al. Endoscopic Raman spectroscopy enables objective diagnosis of dysplasia in Barrett's esophagus. *Gastrointest Endosc* 2014;79:37-45.
18. Bergholt MS, Zheng W, Ho KY, et al. Fiberoptic confocal raman spectroscopy for real-time in vivo diagnosis of dysplasia in Barrett's esophagus. *Gastroenterology* 2014;146:27-32.
19. Bergholt MS, Zheng W, Lin K, et al. Characterizing variability in in vivo Raman spectra of different anatomical locations in the upper gastrointestinal tract toward cancer detection. *J Biomed Opt* 2011;16:037003.
20. Feng S, Zheng Z, Xu Y, et al. A noninvasive cancer detection strategy based on gold nanoparticle surface-enhanced raman spectroscopy of urinary modified nucleosides isolated by affinity chromatography. *Biosens Bioelectron* 2017;91:616-22.
21. Ishigaki M, Maeda Y, Taketani A, et al. Diagnosis of early-stage esophageal cancer by Raman spectroscopy and chemometric techniques. *Analyst* 2016;141:1027-33.
22. Wang J, Lin K, Zheng W, et al. Simultaneous fingerprint and high-wavenumber fiber-optic Raman spectroscopy improves in vivo diagnosis of esophageal squamous cell carcinoma at endoscopy. *Sci Rep* 2015;5:12957.
23. Bergholt MS, Zheng W, Lin K, et al. In vivo diagnosis of esophageal cancer using image-guided Raman endoscopy and biomolecular modeling. *Technol Cancer Res Treat* 2011;10:103-12.
24. Zhou X, Chen GY, Zhang JM, et al. [Research on early diagnosis of esophageal cancer by Raman spectroscopy of human hemoglobin]. *Guang Pu Xue Yu Guang Pu Fen Xi*

- 2013;33:2989-92.
25. Yu B, Cao C, Li P, et al. Sensitive and simple determination of zwitterionic morphine in human urine based on liquid-liquid micro-extraction coupled with surface-enhanced Raman spectroscopy. *Talanta* 2018;186:427-32.
 26. Zhang W, Xia W, Lv Z, et al. Liquid Biopsy for Cancer: Circulating Tumor Cells, Circulating Free DNA or Exosomes? *Cell Physiol Biochem* 2017;41:755-68.
 27. Sharma N, Takeshita N, Ho KY. Raman Spectroscopy for the Endoscopic Diagnosis of Esophageal, Gastric, and Colonic Diseases. *Clin Endosc* 2016;49:404-7.
 28. Buckley K, Ryder AG. Applications of Raman Spectroscopy in Biopharmaceutical Manufacturing: A Short Review. *Appl Spectrosc* 2017;71:1085-116.

Cite this article as: Hao J, Chen C, Jin H, Chen N, Zhou J, Zhu Y, Chung K, Pu Q. The efficacy of Raman spectroscopy in the diagnosis of esophageal cancer: a systematic review and meta-analysis. *Transl Cancer Res* 2020;9(8):4750-4761. doi: 10.21037/tcr-20-854