



Bioinformatics-based identification of differentiated expressed microRNA in esophageal squamous cell carcinoma

King Wai Lau^{1#}, Haikang Zeng^{2#}, Hengrui Liang^{2#}, Xi Su², Jieling Ma², Shuai Wen², Jin Li²

¹Wolfson Institute of Preventive Medicine, Queen Mary University of London, London, UK; ²State Key Laboratory of Respiratory Disease, The First Affiliated Hospital of Guangzhou Medical University, National Clinical Research Center for Respiratory Disease, Guangzhou 510120, China

Contributions: (I) Conception and design: KW Lau, J Li, H Zeng, H Liang; (II) Administrative support: J Li; (III) Provision of study materials or patients: H Zeng, H Liang; (IV) Collection and assembly of data: H Zeng, H Liang; (V) Data analysis and interpretation: X Su, J Ma, S Wen; (VI) Manuscript writing: All authors; (VII) Final approval of manuscript: All authors.

[#]These authors contributed equally to this work.

Correspondence to: Jin Li. State Key Laboratory of Respiratory Disease, The First Affiliated Hospital of Guangzhou Medical University, National Clinical Research Center for Respiratory Disease, Guangzhou 510120, China. Email: lij1250@hotmail.com.

Background: Although numerous studies have identified and observed altered expression of microRNAs in esophageal squamous cell carcinoma (ESCC), only a limited number of miRNAs were reported up to date, partially due to the limitation of sample size (less than or equal to 100 pair of paired samples). Thus, we performed a comprehensive analysis to improve the ability to detect miRNA expressed differentially in ESCC.

Methods: The study datasets were systematically searched and downloaded from public available databases including European Bioinformatics Institute (EMBL-EBI), ArrayExpress and Gene Expression Omnibus (GEO) database. Only datasets derived from ESCC patients were further screened and quality assessed using R programming language with ArrayExpress package. A total of 4 datasets covering 349 ESCC cases and 326 normal esophageal tissue samples (NE) were included in this study.

Results: The analytic results showed that a total of 108 miRNAs were differentially expressed in esophageal cancer, of which 48 were up-regulated and 60 were down-regulated compared with the adjacent normal esophageal tissues. Moreover, we successfully identified 9 novel differentially expressed miRNAs that have not been discovered to associate with esophageal cancer in the previous studies. We also predicted top 5 potential target genes of these novel miRNAs.

Conclusions: The bioinformatics based analysis summarized the current differential expression of miRNA in ESCC, and exploring unknown miRNA target genes provides guidance for discovering the new biomarkers of ESCC.

Keywords: MicroRNAs; esophageal squamous cell carcinoma (ESCC); target gene; GEO database; European Bioinformatics Institute (EMBL-EBI); ArrayExpress

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Introduction

Esophageal squamous cell carcinoma (ESCC), accounts for approximately 90% of cases of esophageal cancer worldwide (1), with 5-year survival rate declining from 20% to 4% in the advanced cases (2). Such a high mortality

rate is partially attributed to late diagnosis, few therapeutic options and poor response to treatment (3,4). Therefore, the identification of new biomarkers for the early detection of ESCC and development of new target-directed therapies are urgently needed.

MicroRNA (miRNAs) is a class of small, highly conserved

noncoding RNAs that negatively regulate gene expression through mRNA degradation or translational inhibition via 3'-untranslated region (3'-UTR) of the modulated gene (5). A given miRNA may have one or multiple target genes and a given gene may be regulated by several miRNAs of the same or different sequence. These short nucleotides can also be over or under expressed, promoting or impeding tumor formation based on the downstream target genes of miRNAs controlled. The miRNA expression profiles of esophageal cancer are significantly different from that of adjacent normal tissue. Several studies have demonstrated that miRNA is related to both the formation and development of EC (6). Hsa-miR-21 is found to be upregulated in squamous cell carcinoma tumor tissues and play a role in abnormal proliferation (7). Hsa-miR-203 is one of the down-regulated miRNAs in esophageal cancer and mediates the post-transcriptional suppression of SOCS3 (8). Analysis of miRNAs also has strong potential for the identification of novel prognostic or predictive biomarkers. For instance, miRNA profiles could use as the prediction of resistance to neoadjuvant radio-chemotherapy in squamous cell carcinoma of the esophagus, which indicated the miRNA are involved in the therapeutic response in EC and suggest that miRNA profiles could facilitate pre-therapeutic patient selection (9).

Although exhaustive miRNA profiling was performed, only a limited number of esophageal cancer associated miRNAs were reported (10-14). All these studies were restricted by the number of clinical samples available (less or equal to 100 tumors and matched nearby normal tissue samples). Thus, to overcome this limitation and to provide a comprehensive landscape of microRNA expression in esophageal cancer, we carried out a pooled-analysis using four public available datasets (12,15-17) comprising 675 patient samples including 349 ESCC cases and 326 normal esophageal tissues (NE). Our analysis discovers 9 new miRNAs. We add the rank of two miRNA to gene prediction to find the target genes.

Methods

Database search and collection strategy

Four datasets E-GEOD-13937, E-GEOD-43732, E-GEOD-6188 and E-GEOD-55856 were downloaded from The European Bioinformatics Institute (EMBL-EBI), ArrayExpress database (<https://www.ebi.ac.uk/arrayexpress/>), according to following criteria: (I) the

number of patients in each study is greater than 50; (II) tumor should company with corresponding normal esophageal tissue from the same patient; (III) all raw data must be available for download and process prior to further analysis; (IV) to reduce bias, dataset generated from the same detection platform was preferred; (V) each patient sample is unique and no duplicated samples were allowed in the analysis.

Annotation

The latest version of human miRNA database (release 21) is downloaded from the mirBase website (18) (<http://www.mirbase.org/>). A modification was also performed to create a new annotation reference by removing the hyphens in the original miRNA name, i.e., hsa-miR-6859-5p to hsa-mir68595p. For each dataset, the miRNA name is extracted from the annotation file, if necessary, it will be modified: (I) p1 occurs at the end of the miRNA name, then it is substituted by p; (II) the character "*" occurs at the end of the miRNA name, then it is replaced by -3p. Each modification is then modified again and match with the miRNA name referred to. If matched with more than one entry of the reference, the miRNA name is removed. The match numbers of miRNA for each dataset is summarized in *Table 1*.

Statistical analysis

Statistical analysis is performed by R (version 3.4.3) software (19). An additional software package (ArrayExpress) was downloaded from Bioconductor (20). The within dataset normalization is carried out using Robust Multi-array Average (RMA) (21) for two-color microarray and Variance Stabilizing Normalization (VSN) (22) for single color microarray as suggested by ArrayExpress. The normalization method for each dataset is summarized in *Table 1*.

Each miRNA can be included in one or more studies. For each miRNA, we find out which studies have this miRNA, e.g., hsa-mir-617 and hsa-mir-1180 can be found in study 2 and 4. Then we group the miRNA according to the combination of studies, e.g., hsa-mir-617 and hsa-mir-1180 is in the same study group (2,4). If the study group has only one study, it will apply to surrogate variable analysis (SVA) by 3 surrogate variables, representing age, smoking and alcohol consumption. If the study group has more than one study, the miRNAs is normalized by Combat (23) to adjust the unknown batch effect, and then further

Table 1 Study characteristics

Study No.	Released date	ArrayExpress	Publication title	NCS	NTS	NUMM	TMUM	NM
1	09-Jul-10	E-GEOD-13937	MicroRNA expression in squamous cell carcinoma and adenocarcinoma of the esophagus: associations with survival	44	44	357	479	VSN
2	01-Aug-14	E-GEOD-43732	MiRNA expression profile study reveals a prognostic signature for esophageal squamous cell carcinoma	119	119	1,719	1887	VSN
3	30-Oct-06	E-GEOD-6188	Distinctive microRNA profiles relating to patient survival in esophageal squamous cell carcinoma	78	55	237	313	VSN
4	30-Jun-16	E-GEOD-55856	Integrated genomic analysis of recurrence-associated small non-coding RNAs in esophageal cancer	108	108	876	1078	RMA

NCS, number of case samples; NTS, number of control samples; NUMM, number of unique mapped miRNA; TMUM, total number of unique miRNA; NM, normalization method; VSN, Variance Stabilizing Normalization; RMA, Robust Multi-array Average.

adjusted using SVA by 3 surrogate variables. The association is calculated using one-way ANOVA, where the dependent variable is the normalized expression level of one miRNA and explanatory variables is a binary variable, where 0 is control and 1 is ESCC case. The ranked-ANOVA is also used, and the mean difference of rank between ANOVA and ranked-ANOVA is smaller than 3 among the top 50 ranked miRNAs. ANOVA has an advantage of estimating the effect size, and hence chosen in the final analysis.

Results

Study characteristics

Four miRNA microarray datasets with more than 300 patient samples were selected for this study (Table 1). All samples are matched tumor and normal. A small subset of data in both E-GEOD-13937 and E-GEOD-6188 dataset was derived from esophageal adenocarcinoma (EAC) and was excluded from this study. Only data collected from ESCC cancers and the adjacent normal tissues were subjected to further investigation. A total of 349 ESCC cases and 326 normal esophageal tissue samples (NE) were covered in this pooled analysis.

Known differentially expressed miRNAs in ESCC

Several recent lines of evidence suggested that miRNAs expression profiling could serve as both diagnostic and prognostic biomarkers in ESCC. We found 20 previously

discovered miRNA in ESCC as known miRNAs. Figure 1 shows the volcano plot of the known differentially expressed miRNAs, and these miRNAs are roughly symmetrical distributed according to the positive or negative effect value. A summary table of the known miRNAs can be found in Table 2. There are 10 miRNAs with more abundant expression in adjacent normal tissues than in ESCC tissues, while there are 10 miRNAs with smaller expression in adjacent normal tissues than in ESCC tissues.

Upregulated miRNAs in ESCC

Many oncogenic miRNAs have been found to be upregulated in our study, e.g., hsa-mir-503, hsa-mir-21-5p, hsa-mir-1290, hsa-mir-1246. Among them, hsa-mir-503 is the most significant upregulated miRNA whose expression was knowingly downregulated in esophagus carcinoma cells (24). The second significantly upregulated one is hsa-mir-21-5p and the one ranking fifth is another mature body of hsa-mir-21, the hsa-mir-21-3p. Previous studies have also confirmed that hsa-mir-21 is predominantly up-regulated in ESCC (25). The third significantly upregulated miRNA is hsa-mir-1290, which has been confirmed expression upregulated in ESCC (26).

Downregulated miRNAs in ESCC

Table 2 also shows some of the miRNAs which were significantly downregulated in ESCC. Among them, hsa-

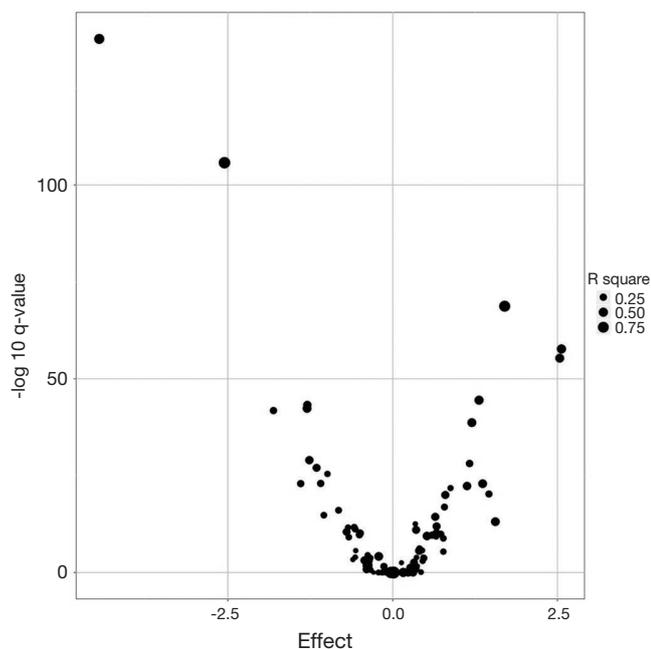


Figure 1 Volcano plot of known differential miRNA expression. Plots were constructed using Effect and $-\log_{10}q$ -value. The size of each dot represents the different R square value.

mir-375 was the most significantly downregulated miRNA confirmed by previous studies (27). The second and third significantly down-regulated miRNAs are hsa-mir-139 and hsa-mir-30a respectively, and both miRNA are confirmed by previous studies in ESCC (10,28).

Previously unknown differentially expressed miRNAs in ESCC

MiRNA not mentioned as significant in published paper is considered as unknown miRNAs. It is because of these miRNAs are tested but not known as associated with ESCC. Table 3 shows the top 9 significantly unknown differentially expressed miRNAs. Only one of them was significantly down-regulated while the rest were significantly up-regulated. The first and second significantly up-regulated unknown miRNAs are hsa-mir-1180 and hsa-mir-663b respectively. The only down-regulated unknown miRNA was hsa-mir-617, which none of the studies mentioned significantly downregulated expressed in ESCC.

With the top 3 predicted target genes for each unknown differentially expressed miRNAs, we apply TopGo to find that GO term positive regulation of cell proliferation (GO:0008284) is the most significant enriched GO term (Table 4).

MiRNA target prediction strategy

The analytical strategy to predict the biological targets of ESCC associated miRNAs is to average the ranked output data from three popular miRNA database, TargetScan (29), Diana (30) and mirDB (31), followed by sorting them in ascending order. Top five target genes for each miRNA are listed in Table 3.

Discussion

Recently, a large number of differentially expressed miRNAs have been observed in esophageal cancer, which supported the value of miRNAs in diagnosis and prognosis. Some studies used qRT-PCR to study a limited number of miRNAs with up to 100 matched tumor and normal samples (7,11), and other experiments used microarrays and qRT-PCR (12-14). In our study, the combination of these four miRNAs microarrays included up to 675 samples [349 ESCC cases and 326 normal esophageal tissue samples (NE)], of which the sample size and number of patients both increased. To our knowledge, this is the most comprehensive meta-analysis to date. For known miRNAs, 95 out of 108 miRNAs can be found in more than one study.

The most significantly upregulated known miRNA was hsa-mir-503. Interestingly, many studies showed that it is down-regulated in many cancers compared with adjacent normal tissues, such as cervical cancer, NSCLC, osteosarcoma, EEC, cervical cancer and hepatocellular carcinoma (32-36). Another study also found the downregulation of miR-503 expression in ESCC tissue relative to adjacent normal tissues in ESCC (37). However our comprehensive pooled analysis of four microarrays indicated that hsa-mir-503 is an upregulated miRNA. Target gene prediction showed that the most promising targets are CCNE1 and FGF2. CCNE1 (also known as Cyclin E1) is a well-known gene associated with cell proliferation and FGF2 is a frequently mutated cancer gene in ESCC (38).

A recent publication shows that hsa-mir-503 downregulated HOXC13, by directly targeting its 3'-UTR, and inhibited proliferation of ESCC. Previous studies have also confirmed that hsa-mir-21, the second significant differentiated miRNA, is predominantly up-regulated in ESCC (28) and has shown it significantly increased in heavy drinking patients. The third significantly upregulated miRNA is hsa-mir-1290, which has been shown that the targeting site of hsa-mir-1290 is located in 3'-UTR of

Table 2 Previously discovered miRNA associated with ESCC

Name	Effect	P value	R square	No. of sample	No. of study	Studies	Known differentially miRNA
hsa-mir-503	1.9757	7.86E-114	0.5407	675	4	1, 2, 3, 4	ESCC > NE
hsa-miR-21-5p	1.6986	8.44E-73	0.772	238	1	4	ESCC > NE
hsa-mir-1290	2.5621	9.85E-62	0.4893	454	2	2, 4	ESCC > NE
hsa-mir-1246	2.5342	2.49E-59	0.46	454	2	2, 4	ESCC > NE
hsa-mir-130b	1.3103	1.62E-48	0.4834	437	3	1, 2, 3	ESCC > NE
hsa-miR-21-3p	1.2024	1.06E-42	0.449	326	2	3, 4	ESCC > NE
hsa-mir-424	1.1657	3.57E-32	0.2768	437	3	1, 2, 3	ESCC > NE
hsa-mir-183	1.3653	5.95E-27	0.4294	437	3	1, 2, 3	ESCC > NE
hsa-mir-34c	1.4591	2.93E-24	0.2476	437	3	1, 2, 3	ESCC > NE
hsa-mir-93	0.7981	5.12E-24	0.3549	437	3	1, 2, 3	ESCC > NE
hsa-mir-375	-4.451	8.64E-142	0.6188	675	4	1, 2, 3, 4	ESCC < NE
hsa-miR-139-5p	-2.5485	8.54E-110	0.8855	238	1	4	ESCC < NE
hsa-miR-30a-3p	-1.2934	2.91E-47	0.3731	459	3	1, 3, 4	ESCC < NE
hsa-mir-133b	-1.8072	8.00E-46	0.2604	675	4	1, 2, 3, 4	ESCC < NE
hsa-mir-29c	-1.2628	5.34E-33	0.3599	437	3	1, 2, 3	ESCC < NE
hsa-mir-99a	-1.1523	5.19E-31	0.3341	437	3	1, 2, 3	ESCC < NE
hsa-mir-100	-1.0906	5.50E-27	0.2541	437	3	1, 2, 3	ESCC < NE
hsa-mir-139	-1.394	6.22E-27	0.2645	437	3	1, 2, 3	ESCC < NE
hsa-mir-150	-1.0424	9.39E-19	0.2014	437	3	1, 2, 3	ESCC < NE
hsa-mir-625	-1.5571	4.06E-17	0.4263	216	1	2	ESCC < NE

ESCC, esophageal squamous cell carcinoma; NE, normal esophageal.

NFIX (29). Also the inverse correlation between the levels of hsa-mir-1290 and NFIX protein and miRNA in ESCC tissue samples were also confirmed.

Hsa-mir-375 was the most significantly downregulated miRNA confirmed by previous studies (30). It has been shown that SHOX2 is a direct target of hsa-mir-375, where SHOX2 expression is upregulated in ESCC. Further analysis showed that SHOX2 induced proliferation, invasion, and metastasis of ESCC. The second and third significantly down-regulated miRNAs is hsa-mir-139 and hsa-mir-30a respectively. It has been shown that the target of hsa-mir-139 is NR5A2, which induce cell cycle arrest in the G0/G1 phase and to suppress the invasive capability of esophageal carcinoma cells (10). MiRNA hsa-mir-30a is closely associated with advanced ESCC progression and poor prognosis of patients with ESCC by promoting ESCC cell proliferation (31).

At present, many studies have verified that the tumorigenic with hsa-mir-21 upregulation in ESCC is associated with target genes PDCD4, K-RAS, PTEN and ERK1/2/MAPK pathway (39-42). Meanwhile, a study suggested that hsa-mir-21 down-regulation suppresses cell growth, invasion and induced cell apoptosis by targeting FASL, TIMP3, and RECK genes in esophageal carcinoma (26). Several studies have found that hsa-mir-1290 promotes cancer progression by targeting nuclear factor I/X (NFIX) and SCAI in ESCC (43,44). Similarly, hsa-mir-130b plays a carcinogenic role by inhibiting PTEN expression in ESCC (45). In summary, these markedly upregulated miRNAs are predominantly tumorigenic, and they are likely to be prognostic biomarkers for ESCC *Table 5*.

The most significantly downregulated miRNA in our study is hsa-mir-375. Accumulated evidence indicated that this microRNA rendered its strong tumor-suppressive effect

Table 3 Previously undiscovered miRNA associated with esophageal squamous cell carcinoma (ESCC)

Name	Effect	P value	R square	No. sample	No. study	Studies	Target gene list
hsa-mir-617	-3.3138	1.50E-103	0.6615	454	2	2, 4	AASDH, RSF1, EPHA10, EPC1, STXBP1
hsa-mir-1180	1.6203	2.25E-56	0.5648	454	2	2, 4	CBX5, MAPK1, FOXN1, TFEB, HOXB5
hsa-mir-663b	1.6974	9.61E-55	0.4952	454	2	2, 4	CHTF8, MAP7D1, AC090673.1, DHX30, MAP7D1
hsa-mir-421	1.2688	1.52E-49	0.4871	454	2	2, 4	AGO2, DNAJC27, SLC25A3, ANKRD17, MEF2D
hsa-mir-196b	2.0919	3.71E-49	0.4193	437	3	1, 2, 3	Not applicable
hsa-mir-370	1.1493	1.81E-44	0.2556	675	4	1, 2, 3, 4	SIK3, AGO2, PFKM, ATXN7L3, USP43
hsa-mir-711	0.7607	2.22E-44	0.6275	454	2	2, 4	NSD3, C2CD2, NECAP1, C2CD2, MCIDAS
hsa-mir-2276	0.6688	3.20E-43	0.5666	454	2	2, 4	RNF111, MED1, DST, RGPD1, HNRNPR
hsa-mir-2278	0.7325	8.90E-43	0.4582	454	2	2, 4	PAPSS2, CTNNA1, PAFAH1B1, KDM2A, PAPP

through targeting at SHOX2, MMP13, LDHB, AEG-1/MTDH, IGF1R and PDK1 genes (46-50). hsa-mir-139-5p is one of miRNA previously discovered to suppress tumor growth and invasiveness in human ESCC through unknown biological targets (51) (Table 5). The target of has-mir-139a is NR5A2 and hsa-mir-30a is associated with ESCC cell proliferation.

The most significantly up-regulated miRNA is hsa-mir-1180, which may modulate the expression of predicted target gene MAPK1 and participate in a variety of cellular processes such as proliferation, differentiation, transcription regulation and development. Surprisingly, the down-regulation of this miRNA has been observed in pancreatic cancers with anti-cancer activity via regulation of the TNIP2/NF-KB signaling pathway and its downstream proteins, including MMP-2, MMP-9, Bax, Bcl-2, p21, and cyclin D1 (52).

The most significantly downregulated miRNA was hsa-mir-617, which was one of the top 9 significantly differentially expressed unknown miRNAs. To our knowledge, we are the first research group to demonstrate the reverse association of this miRNA with esophageal cancer. A negative correlation in the expression of this microRNA and tumor has also been observed in oral cavity and oral verrucous carcinoma (53). Five potential modulated genes of this microRNA are AASDH, RSF1, EPHA10, EPC1 and STXBP1. Among these targets, RSF1

was related with tumor proliferation, invasion and chemoresistance, and EPC1 promotes cell apoptosis and DNA repair.

We have to admit limitations in this analysis. Firstly, Clinical characteristics of patients are collected from the GEO and ArrayExpress database. None of clinical characteristics of patients are present in all four studies, and hence we could not use any clinical characteristics of patients. However, we apply Surrogate Variable Analysis (SVA) to take into account most commonly known confounder variables, i.e., age, smoking and alcohol consumption. Secondly, this study focus on ESCC only. In Europe, it is preferably a sporadic disease with a stable incidence, while in some areas, like China, Iran, Pakistan, it is rather an endemic disease with a high prevalence. Thus, our analysis might not represent a genomic map of all race. Third, the “newly described” miRNA in the analysis is based on the time when that four included arrays were conducted. Some might be defined later by other studies but it still showed a “new” one here. Last, ESCC patients enrolled at different clinical centers from the United States, Canada, China and Japan, thus multi-racial study makes it impossible to reveal one racial character, but it would represent more population worldwide.

Since the first ESCC related miRNA was reported in 2006, both the samples size and number of patients in relevant studies are continually increasing. We have

Table 4 Biological process: pathways analysis for previously unknown differentially expressed miRNAs

GO ID	GO term	Annotated	Significant	Expected	Elim	Geneset
GO:0008284	Positive regulation of cell proliferation	866	14	5.57	0.0013	<i>HIPK1; CHTOP; PROX1; ITGB1; NOG; FAM98A; CUL3; DPP4; MAPK1; COX17; PDCD10; WDR48; MYB; PDCD1LG2</i>
GO:0016032	Viral process	775	13	4.99	0.0014	<i>NFIA; PROX1; ITGB1; ZMYND11; RSF1; RAB6A; CBX5; PABPN1; UBN1; DPP4; MAPK1; WDR48; AP1S2</i>
GO:0031062	Positive regulation of histone methylation	36	3	0.23	0.0016	<i>CHTOP; MYB; PHF19</i>
GO:0050673	Epithelial cell proliferation	360	8	2.32	0.0023	<i>PROX1; BCL11B; NOG; FOXN1; MAPK1; PDCD10; WDR48; PTCH1</i>
GO:0021877	Forebrain neuron fate commitment	12	2	0.08	0.0026	<i>SMARCC2; BCL11B</i>
GO:0033631	Cell-cell adhesion mediated by integrin	12	2	0.08	0.0026	<i>ITGB1; DPP4</i>
GO:0034063	Stress granule assembly	13	2	0.08	0.0031	<i>ATXN2; C9orf72</i>
GO:0035278	MiRNA mediated inhibition of translation	13	2	0.08	0.0031	<i>TNRC6A; AGO2</i>
GO:0048538	Thymus development	46	3	0.3	0.0032	<i>BCL11B; FOXN1; MAPK1</i>
GO:0010468	Regulation of gene expression	4585	48	29.49	0.0043	<i>HIPK1; CHTOP; NFIA; PROX1; LCOR; ZMYND11; RSF1; SF1; EMSY; PIDD1; HIPK3; FLI1; TNKS1BP1; ATXN2; CBX5; SARNP; SMARCC2; PFKM; PABPN1; BCL11B; TLE3; UBN1; TNRC6A; ZNF267; TERF2; ZFP3; NOG; FOXN1; ZBTB4; FAM98A; CUL3; NRIP1; MAPK1; KBTBD8; MBNL1; PDCD10; SETD5; DPH3; ZNF827; DAB2; MYB; ZNF800; CREB5; AGO2; PTCH1; PHF19; RC3H2; ZMYM3</i>

Table 5 Previously studies about top 5 known miRNAs mechanism associated with esophageal squamous cell carcinoma (ESCC)

Known miRNAs	Regulation in ESCC	Function	Target genes or downstream pathway
hsa-miR-21	↑	Carcinogenesis	<i>PDCD4, K-RAS, PTEN, ERK1/2/MAPK</i>
hsa-miR-21	↓	Anti-carcinoma	<i>FASL, TIMP3, RECK</i>
hsa-mir-1290	↑	Carcinogenesis	<i>NFIX, SCAI</i>
hsa-mir-1246	↑	Carcinogenesis	Not applicable
hsa-mir-130b	↑	Carcinogenesis	<i>PTEN</i>
hsa-mir-375	↓	Anti-carcinoma	<i>MTD, SHOX2, MMP13, LDHB, AEG-1/MTDH, IGF1R, PDK1</i>
hsa-mir-139	↓	Anti-carcinoma	Not applicable
hsa-mir-30a	↓	Anti-carcinoma	Not applicable
hsa-mir-133b	↓	Anti-carcinoma	<i>FSCN1, SQLE</i>
hsa-mir-29c	↓	Anti-carcinoma	Cell cycle G(1)/G(0)

performed the largest meta-analysis study with individual patient data on ESCC miRNA microarrays, which provided guidance for discovering the new biomarkers of ESCC for early diagnosis and targeted therapies. The role and mechanism of these miRNAs in ESCC need to be further investigated.

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Footnote

Conflicts of Interest: All authors have completed the ICMJE uniform disclosure form (available at <http://dx.doi.org/10.21037/tcr.2018.10.15>). The authors have no conflicts of interest to declare.

Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. The study was conducted in accordance with the Declaration of Helsinki (as revised in 2013). Institutional ethical approval and informed consent were waived.

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