Impact of apatinib in combination with osimertinib on EGFR T790M-positive lung adenocarcinoma

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Background: The purpose of this study was to investigate the anti-tumor activities and the mechanisms of the third-generation epidermal growth factor receptor (EGFR) tyrosine kinase inhibitor (TKI) osimertinib, combined with the anti-angiogenic target drug apatinib, in the treatment of lung adenocarcinoma. We investigated the effects of these drugs in vitro in PC9 (E19 del) and H1975 (E21 L858R/E20 T790M) cell lines, as well as in vivo in both mouse and human experiments.

Methods: PC9 and H1975 cells were cultured in 96-well plates and incubated with osimertinib (1–100 nmol/L), or apatinib (100–1,000 nmol/L), or a combination of the two agents, for 48 h. Cell viability was determined using a Cell Counting Kit-8. The protein expression of EGFR and its downstream signaling pathway members (AKT and ERK) was detected by western blot. For in vivo experiments, BALB/c nude mice were subcutaneously inoculated with H1975 cells in a xenograft model of adenocarcinoma. Mice bearing tumors were treated with osimertinib alone or in combination with apatinib, and tumor growth curves were obtained. Furthermore, we evaluated the efficacy and safety of combined osimertinib and apatinib therapy in three patients with EGFR T790M positive lung adenocarcinoma, who had been previously sensitized to osimertinib but developed an acquired resistance.

Results: In vitro experiments revealed that osimertinib combined with apatinib increased the growth inhibition of PC9 and H1975 cells, simultaneously reducing the protein expression of phosphorylated EGFR and its downstream signaling pathway members in H1975 cells, compared to osimertinib treatment alone. In vivo experiments revealed that the combination of osimertinib and apatinib decreased tumor volume in an H1975 cell xenograft model, compared to osimertinib monotherapy at different dosages. All three patients with T790M positive lung adenocarcinoma that progressed following osimertinib treatment responded to continuous osimertinib in combination with apatinib, with a progression-free survival (PFS) range of 5–7 months.

Conclusions: Apatinib can enhance the anti-tumor activity of osimertinib in the treatment of T790M positive lung adenocarcinoma. Further clinical studies are needed to confirm these results.

Keywords: Non-small cell lung cancer (NSCLC); epidermal growth factor receptor (EGFR); tyrosine kinase inhibitor (TKI); osimertinib; apatinib

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Introduction

Epidermal growth factor receptor (EGFR) tyrosine kinase inhibitors (TKIs) are the standard first-line therapy for patients with EGFR mutated advanced non-small cell lung cancer (NSCLC). First- or second-generation EGFR-TKIs can improve the objective response rate (ORR) by 60–80%, as well as the median progression-free survival (PFS) by 9–13 months (1-3). While most patients’ cancer will eventually progress due to secondary resistance, approximately 50–60% of cases are mediated by an acquired T790M mutation (4,5). Osimertinib is a so-called third-generation EGFR-TKI, targeting both EGFR sensitive mutations and the T790M mutation. A phase III study, AURA-3, indicated that osimertinib resulted in an ORR of 71% and a median PFS of 10.1 months for advanced NSCLC patients, who acquired the T790M mutation after resistance to first-line EGFR-TKIs (6). The resistance mechanism of osimertinib is more complex compared to first- or second-generation EGFR-TKIs. Currently, there are no effective targeted drugs to treat patients who have become resistant to osimertinib, with chemotherapy being the standard route of care.

Previous studies have shown that bevacizumab, an anti-vascular endothelial growth factor (VEGF) antibody, can enhance anti-tumor activity of chemotherapy and delay the onset of drug resistance (7). Preclinical studies have confirmed that EGFR-TKIs combined with bevacizumab can partially reverse the resistance to EGFR-TKIs (8,9). Subsequently, a phase II clinical study has shown that EGFR-TKI combined with bevacizumab treatment results in a significantly prolonged PFS compared to EGFR-TKI alone (10). The reason for this may be that the EGFR and VEGF/VEGFR signaling pathways are tightly interconnected. EGFR-TKIs increase VEGF levels in tumors, which consequently negatively regulate the tumor vascular microenvironment. However, anti-angiogenic drugs can lower VEGF levels and angiogenesis, promote tumor vessel normalization, improve tumor oxygenation, decrease interstitial fluid pressure, and improve drug delivery to tumors, thereby inducing an increased sensitivity of the tumor cells to EGFR-TKIs (11).

Apatinib [AiTan™ (China); Rivoceranib® (global)] is a novel small molecule that is a selective VEGFR-2 TKI, which has been approved by the China Food and Drug Administration for the third-line treatment of advanced gastric cancer (12,13). It can also effectively inhibit the activity of Ret, c-kit, c-src, and platelet-derived growth factor receptor β (PDGFR-β), thus promoting anti-tumor activity (14). A phase II study indicated that apatinib had promising anti-tumor activity in advanced NSCLC, with lower and more manageable grade 3/4 toxicity events (15). A Phase III clinical study comparing the efficacy and safety of apatinib vs. placebo as a third-line treatment for EGFR mutation-negative advanced NSCLC is ongoing, and the results have yet to be reported.

Due to the multi-targeted anti-tumor effect and acceptable safety profile of apatinib, we aimed in the present study to investigate whether the combined therapy of a third-generation EGFR-TKI, namely osimertinib and apatinib, could further improve the anti-tumor activity and delay the occurrence of acquired resistance for lung adenocarcinoma harboring the acquired EGFR T790M mutation, compared to single EGFR-TKI treatment alone.

Methods

Cell culture and reagents

The human NSCLC cell lines PC9 (exon 19 deletion) and H1975 (exon 21 L858R and exon 20 T790M) were obtained from the American Type Culture Collection. Cells were cultured in Dulbecco’s modified Eagle’s medium (DMEM) media (Biological Industries, Kibbutz Beit Haemek, Israel) or RPMI 1640 media (Biological Industries), supplemented with 10% fetal bovine serum (FBS) and 1% penicillin-streptomycin at 37 °C in 5% carbon dioxide. Osimertinib and apatinib were purchased from Selleck Chemicals (Shanghai, China).

Cell growth assay

PC9 and H1975 cell lines were seeded at a density of 2,000–5,000 cells per well in 96-well plates. After 24 h of incubation, cells were exposed to osimertinib (1–100 nmol/L), apatinib (100–1,000 nmol/L), or a combination of the two agents for 48 h in complete media. Cell viability was determined using a Cell Counting Kit-8 (Biological Industries). The absorbance was measured at 450 nm based on the principle of water-soluble formazan dye development from tetrazolium salt. The dose administered was based on the results of the preliminary experiment and data from other articles. Data are shown as the mean ± standard deviation (SD) of at least three independent experiments performed in triplicate.
**Western blotting**

Cells (H1975) were seeded into 6-well plates at a concentration of 5×10^5 cells per well, and were incubated overnight. Then, osimertinib, apatinib or a combination of the two agents were added, and cells were incubated continuously for 48 h. Cells were lysed in radio immunoprecipitation assay (RIPA) buffer containing phenylmethanesulfonyl fluoride (PMSF) with mild sonication. The total protein concentration was measured using a BCA Protein Assay Kit (Beyotime Biotechnology, Beijing, China). Equal amounts of protein (40–50 mg) were subjected to sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) using 10% Bis-Tris precast gels (Beyotime Biotechnology), and transferred onto polyvinylidene difluoride (PVDF) membranes (Millipore, Hayward, CA, USA) using the semi-dry transfer system. The membranes were blocked in 5% bovine serum albumin-tris-buffered saline with Tween (BSA-TBST) or milk-TBST for 1 h at room temperature, and blotted with primary antibodies against total EGFR, phospho-EGFR, total AKT, phospho-AKT, total ERK, phospho-ERK, or GAPDH at 4 °C overnight. Membranes were then incubated with horseradish peroxidase (HRP)-conjugated goat anti-rabbit secondary antibodies or anti-mouse secondary antibodies for 2 h at room temperature, and visualized by electrochemiluminescence (ECL). The image was scanned and analyzed using Image J (National Institutes of Health, Bethesda, MA, USA), and protein levels were normalized to the level of GAPDH.

**Animals and tumor xenograft experiments**

Five to 6-week-old female BALB/c nude mice were purchased from Beijing Huafukang Biotechnology Co. (Beijing, China). Mice were provided with food and water, and housed under a 12 h light-dark cycle. Mice were allowed to acclimatize for 1 week before any handling. The experimental protocols were reviewed and approved by the China Medical University Animal Care and Use Committee (Shenyang, China). The study was conducted by the National Institutes of Health “Principles of Laboratory Animal Care” (NIH publication No. 85-23, revised 1985) guidelines for the care and use of laboratory animals.

H1975 cells were harvested and resuspended in a 4:1 mixture of serum-free RPMI 1640 and Matrigel. Cells ([5–10]×10^6) were injected subcutaneously into the rear flank of mice using a 20-gauge needle. After two weeks, when tumors had grown to 400–600 mm^3, mice were randomized into three groups for dosing: a low dose of osimertinib (n=4, 2.5 mg/kg/day, p.o. gavage; group A), a high dose of osimertinib (n=4, 5 mg/kg/day, p.o. gavage; group B) and a low dose of osimertinib plus apatinib (n=4, osimertinib 2.5 mg/kg/day, p.o. gavage; apatinib 2.5 mg/kg, twice weekly, p.o. gavage; group C). The dose administered was based on the results of the preliminary experiment and data from other articles. The tumor volumes were measured using a digital caliper twice weekly, and were calculated using the following equation: volume (V) (mm^3) = length × width^2 × π/6. Moribund animals were killed to reduce suffering. After 16 days of treatment, mice were euthanized.

**Patients**

The efficacy and safety of combined therapy of osimertinib and apatinib was evaluated in eligible patients with EGFR T790M positive lung adenocarcinoma. Eligible patients were previously sensitized to osimertinib, but developed acquired resistance. The enrolled patients were treated with continuous osimertinib (80 mg/day) plus apatinib (500 mg/day) in the hospital, until the disease progressed according to the Response Evaluation Criteria in Solid Tumor Standard (version 1.1), or until unacceptable toxicities developed. All patients were followed up once every 2 months for a chest and abdominal CT scan. All patients provided written informed consent prior to participating in the study. This study was approved by the ethics committee of our hospital, and the study was conducted in accordance with the Declaration of Helsinki.

**Statistical analysis**

All data were analyzed using SPSS software, version 19.0 (SPSS, Chicago, IL, USA). Quantitative data are expressed as the mean ± SD. A Student’s t-test was used for comparisons between two groups. A P value less than 0.05 was considered statistically significant.

**Results**

**Inhibitory effect of osimertinib combined with apatinib on PC9 and H1975 cells in vitro**

The inhibition rate of PC9 and H1975 cells in each group is shown in Figure 1A,B,C,D. PC9 and H1975 lung adenocarcinoma cells were sensitized to osimertinib or
osimertinib combined with apatinib in a dose-dependent manner. The inhibitory rate of PC9 cells was significantly higher than that of H1975 cells, at all drug concentrations (P<0.01). Osimertinib combined with apatinib increased the tumor inhibition rate of PC9 and H1975 cell lines, compared to single osimertinib treatment at the same concentration (P<0.05). Under a fixed concentration of osimertinib, the tumor inhibition rate of PC9 and H1975 cells also increased to a certain extent with increased concentration of apatinib (P<0.05). Additionally, the tumor inhibition rates under low concentrations of osimertinib combined with a high concentration of apatinib (1,000 nmol/L) were similar to that of a high concentration of osimertinib alone (P>0.05).

EGFR and its downstream signaling protein expression in H1975 cells

Western blot results are shown in Figure 2. The protein expression of p-EGFR, p-AKT, and p-ERK gradually decreased with increased osimertinib concentration. Additionally, the expression levels of p-EGFR, p-AKT, and p-ERK in the osimertinib combined with apatinib group were modestly decreased, compared to treatment with apatinib or osimertinib alone (P<0.05). Apatinib monotherapy resulted in negligible inhibition of protein expression. Total EGFR, total AKT, and total ERK protein expression was unchanged in each group.

Anti-tumor activity of osimertinib combined with apatinib in xenograft models

The tumor growth in each group of mice after two weeks of treatment are shown in Figure 3A,B,C. Osimertinib monotherapy or a combination of osimertinib and apatinib resulted in tumor inhibition and a reduced tumor volume of H1975 lung adenocarcinoma cell xenograft models, to differing levels. Specifically, tumor volume was more highly
Figure 2 Effect of osimertinib and apatinib on EGFR and its downstream signaling pathway protein expression in H1975 cells. The units of osimertinib and apatinib are nmol/L. Osi, osimertinib; Apa, apatinib; EGFR, epidermal growth factor receptor.

Figure 3 Results of osimertinib monotherapy vs. combination of osimertinib and apatinib in H1975 cell xenograft models. (A) Images of nude mice in each group. Mice were treated on day 14 with osimertinib alone at 2.5 mg/kg/day, A group; osimertinib alone at 2.5 mg/kg/day, B group; osimertinib 2.5 mg/kg/day and apatinib 2.5 mg/kg, twice weekly, C group. After 16 days of treatment, mice were euthanized. (B) Tumor volume growth curve. Each point represents the mean ± standard deviation of tumor volumes from four mice in each group. The black arrow indicates that the mice were randomly intervened with drugs on the 14th day after inoculation. (C) Images of tumor samples in each group. The samples were taken on day 30 after implantation.
Table 1 Characteristics of three advanced lung adenocarcinoma patients administered therapy

<table>
<thead>
<tr>
<th>Characteristics</th>
<th>Patient #1</th>
<th>Patient #2</th>
<th>Patient #3</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age, years</td>
<td>51</td>
<td>55</td>
<td>53</td>
</tr>
<tr>
<td>Sex</td>
<td>Male</td>
<td>Male</td>
<td>Male</td>
</tr>
<tr>
<td>Stage</td>
<td>IV (T2N0M1)</td>
<td>IV (T4N2M1)</td>
<td>II (T2N1M0)</td>
</tr>
<tr>
<td>EGFR mutation</td>
<td>Exon21 L858R</td>
<td>Exon19 del</td>
<td>Exon19 del</td>
</tr>
<tr>
<td>Acquired T790M mutation</td>
<td>Yes</td>
<td>Yes</td>
<td>Yes</td>
</tr>
<tr>
<td>Treatment [duration, months]</td>
<td>Adjuvant therapy</td>
<td>–</td>
<td>DDP+TAX</td>
</tr>
<tr>
<td></td>
<td>First-line</td>
<td>DDP + PEM + PRT</td>
<td>ERL [10]</td>
</tr>
<tr>
<td></td>
<td>Maintenance</td>
<td>PEM then ERL [45]</td>
<td>–</td>
</tr>
<tr>
<td></td>
<td>Fifth-line</td>
<td>–</td>
<td>Osi + Apa [7]</td>
</tr>
<tr>
<td></td>
<td>Sixth-line</td>
<td>–</td>
<td>Bev + DTX + Gem [1]</td>
</tr>
<tr>
<td>OS₁ (months)</td>
<td>65</td>
<td>43+</td>
<td>90+</td>
</tr>
<tr>
<td>OS₂ (months)</td>
<td>6</td>
<td>13+</td>
<td>13+</td>
</tr>
<tr>
<td>Adverse effects</td>
<td>G3 oral mucositis, G3 fatigue</td>
<td>G2 fatigue</td>
<td>G2 oral mucositis, G3 fatigue</td>
</tr>
</tbody>
</table>

OS₁ was defined as the time from the start of first line of treatment to the date of death by any cause; OS₂ was defined as the time from the start of apatinib combined with osimertinib treatment to the date of death by any cause. DDP, cisplatin; PEM, pemetrexed; PRT, palliative radiotherapy; Osi, osimertinib; Apa, apatinib; ERL, erlotinib; AB-TAX, albumin-bound taxol; WBRT, whole-brain radiotherapy; TMZ, temozolomide; DTX, docetaxel; NDP, nedaplatin; Bev, bevacizumab; Gem, gemcitabine; Oxa, oxaliplatin; G2/3, grade 2/3.

Discussion

In clinical practice, osimertinib is commonly used for the treatment of patients who have acquired the EGFR T790M mutation in advanced NSCLC after secondary resistance to prior first- or second-generation EGFR-TKIs. Osimertinib remarkably improves median PFS compared to the first generation of EGFR-TKIs used as first-line treatments for EGFR sensitizing mutation-positive NSCLC (18.9 versus 10.2 months) (16). Nevertheless, patients who initially respond to osimertinib will inevitably develop resistance. Our study shows that osimertinib in combination with apatinib in the treatment of T790M mutation-positive lung adenocarcinoma can increase anti-tumor efficacy, compared to osimertinib monotherapy. It may even overcome the resistance to osimertinib in patients who initially progressed on osimertinib, resulting in an impressive PFS and manageable toxicity profile. Therefore, the effect of delaying and overcoming the occurrence of resistance to
osimertinib with combined osimertinib and apatinib therapy is worthy of further exploration. We strongly recommend that future larger clinical trials in this population be performed.

The EGFR T790M mutation refers to the mutation of threonine (T) to methionine (G) at amino acid position 790 in exon 20 of EGFR. Because methionine occupies more space than threonine, this results in the formation of steric hindrance, changing the affinity of ATP to the EGFR kinase domain. As a result, EGFR-TKI-like small molecule drugs can no longer effectively block EGFR activation signals, and they therefore lose their ability to kill tumor cells (17,18). Osimertinib has a 2-aminopyrimidine structure as a backbone, and binds irreversibly to the C797 amino acid of the EGFR ATP binding site, selected by unsaturated acryloyl chains. It thereby inhibits EGFR-sensitive mutations in T790M-mutant tumor cells (16). Our study indicates that the inhibitory rate of PC9 cells was higher than that of H1975 cells at the same osimertinib concentration, which is consistent with the results of clinical trials. Patients with an EGFR sensitizing mutation who received osimertinib therapy had a significantly longer PFS than patients with acquired T790M mutation-positive NSCLC who received osimertinib therapy (6,14,17,18). Results from one cohort of the AURA I study and the FLAURA phase III trial demonstrated a median PFS of more than 1.5 years for patients with EGFR sensitizing mutation-positive advanced NSCLC who received osimertinib as the first-line treatment (16,19). However, a median PFS of only 10.2 months was seen in the AURA-3 phase III trial, amongst patients who acquired EGFR T790M mutation-positive advanced NSCLC after developing secondary resistance to prior first- or second-generation EGFR-TKI treatment (6).

The possible mechanisms of secondary resistance to osimertinib include a secondary C797S mutation, loss of T790M, and bypass activation, such as the amplification of MET and HER2 (20). In clinical practice, there is currently no standard therapeutic drug for those patients who both fail multi-line chemotherapy and acquire resistance to the
Table 2 List of preclinical trials using dual inhibition of the VEGF/VEGFR and EGFR pathways: A+T therapy mode

<table>
<thead>
<tr>
<th>Trials</th>
<th>Treatment</th>
<th>Study object/cell lines</th>
<th>Significance</th>
</tr>
</thead>
</table>
| Masuda (8) | Bevacizumab | B901L (Del) | 1. Established a treatment model that became refractory to erlotinib and bevacizumab enhanced antitumor activity  
2. Re-induction of VEGF and subsequent VEGF-dependent tumor growth is suggested to be one of the major mechanisms of acquired resistance to erlotinib |
| Naumov (9) | Bevacizumab | A549 (WT) | 1. Erlotinib resistance may be associated with a rise in both tumor cell and host stromal VEGF  
2. Combined blockade of the VEGFR and EGFR pathways can abrogate primary or acquired resistance to EGFR-TKIs |
| Erlotinib | Calu-6 (WT) | H3255 (L858R) | Combined anti-VEGF therapy enhances the antitumor activity of anti-EGFR therapy and/or partially reverse resistance to EGFR-TKI, by increasing EGFR-TKI concentration in specific tumors that express high levels of VEGF protein |
| Gefitinib | Vandetanib | H1975 cells (L858R/T790M) | Erlotinib  
Bevacizumab + erlotinib | H1650 (Del)  
HCC827 (Del) | Bevacizumab | H157/H460/A549 (WT)  
PC9 (Del) | Bevacizumab + erlotinib | 11–18 (L858R)  
H1975 (L858R + T790M) | Li (22) | Combined anti-VEGF therapy enhances the antitumor activity of anti-EGFR therapy and/or partially reverse resistance to EGFR-TKI, by increasing EGFR-TKI concentration in specific tumors that express high levels of VEGF protein |
| Ito (23) | E7820 | A549 (WT) | The combination of E7820 with erlotinib as an alternative strategy to overcome erlotinib resistance in NSCLC by enhancing the anti-angiogenic activity of E7820 |
| Erlotinib | E7820 + erlotinib | H1650 (Del)  
H1975 (L858R/T790M) | Furugaki (24) | The addition of bevacizumab to erlotinib did not enhance antitumor activity in primarily erlotinib-resistant tumors with the T790M mutation as long as their growth remained significantly suppressed by erlotinib (24). Two randomized control studies reported that erlotinib combined with bevacizumab significantly prolonged the median PFS in patients with EGFR sensitizing mutation-positive NSCLC, compared to

VEGF, vascular endothelial growth factor; EGFR, epidermal growth factor receptor; TKIs, tyrosine kinase inhibitors; WT, EGFR wild-type; Del, EGFR exon 19 deletion; NSCLC, non-small-cell lung cancer; E7820, an angiogenesis inhibitor that decreases integrin-α2 expression.

Before osimertinib was approved for clinical use, many experts explored whether the “A+T” (EGFR-TKIs in combination with anti-angiogenetic drugs) therapy mode could enhance anti-tumor activity, or even reverse resistance to first-generation EGFR-TKIs. The preclinical and clinical trials regarding the “A+T” therapy mode are summarized in Tables 2 and 3, respectively. Most preclinical studies show that combined VEGFR/EGFR pathway blockade can abrogate primary or acquired resistance to EGFR inhibitors (8,9,22,23). Of course, not all preclinical studies support the effectiveness of EGFR-TKIs combined with anti-angiogenic drugs against EGFR-TKI-resistant NSCLC cells. Furugaki et al. indicated that the addition of bevacizumab to erlotinib was ineffective against NSCLC cells or xenograft models that were primarily resistant to erlotinib; however, the combination treatment enhanced the inhibitory effect on T790M mutation- or MET amplification- positive NSCLC cells harboring an EGFR sensitizing mutation, as long as their growth remained significantly suppressed by erlotinib (24). Two randomized control studies reported that erlotinib combined with bevacizumab significantly prolonged the median PFS in patients with EGFR sensitizing mutation-positive NSCLC, compared to
Table 3 List of clinical trials investigating the dual inhibition of the VEGF/VEGFR and EGFR pathways in patients with NSCLC

<table>
<thead>
<tr>
<th>Trials</th>
<th>Patients number</th>
<th>Line of treatment</th>
<th>EGFR mutation status</th>
<th>Treatment region</th>
<th>ORR, % (P value)</th>
<th>mPFS, months (P value)</th>
<th>mOS, months (P value)</th>
</tr>
</thead>
<tbody>
<tr>
<td>West (25)</td>
<td>84 BAC</td>
<td>–</td>
<td>Unclear</td>
<td>Bev + Erl</td>
<td>22</td>
<td>5</td>
<td>21</td>
</tr>
<tr>
<td></td>
<td>85 non-smokers</td>
<td></td>
<td></td>
<td></td>
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<td></td>
</tr>
<tr>
<td>Herbst (26)</td>
<td>120</td>
<td>Recurrent/refractory</td>
<td>Unclear</td>
<td>Chemo (n=41) Bev + Chemo (n=40) Bev + Erl (n=39)</td>
<td>12.2 (P=0.019)</td>
<td>3.0 (P=0.018)</td>
<td>8.6 (P=0.406)</td>
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<tr>
<td>Ciuleanu (27)</td>
<td>124</td>
<td>First-line</td>
<td>Unclear</td>
<td>Bev + Chemo (n=61) Bev + Erl (n=63)</td>
<td>23.8 (P=0.19)</td>
<td>18.4 weeks (P=0.018)</td>
<td>16.4 weeks (P=0.406)</td>
</tr>
<tr>
<td>Wang (28)</td>
<td>297</td>
<td>Second-line</td>
<td>Unclear</td>
<td>Bev + Erl + Pan (n=150) Erl + placebo (n=147)</td>
<td>38 (P=0.014)</td>
<td>4.6 (P=0.003)</td>
<td>10.4 (P=0.031)</td>
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<tr>
<td>Herbst (29)</td>
<td>636</td>
<td>Second-line</td>
<td>Unclear</td>
<td>Bev + Erl (n=319) Erl + placebo (n=317)</td>
<td>13 (P=0.014)</td>
<td>3.4 (P=0.003)</td>
<td>9.3 (P=0.031)</td>
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<td>Ninomiya (30)</td>
<td>19</td>
<td>First-line</td>
<td>Positive</td>
<td>Afa + Bev</td>
<td>81</td>
<td>–</td>
<td>–</td>
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<tr>
<td>Gautschi (31)</td>
<td>97</td>
<td>First-line</td>
<td>Positive</td>
<td>Bev + Erl (n=20) WT Bev + Chemo (n=77)</td>
<td>70 (P=0.0005)</td>
<td>14 (P=0.0157)</td>
<td>Not reached (P=0.0005)</td>
</tr>
<tr>
<td>Yoshida and Seto (10,32)</td>
<td>152</td>
<td>First-line</td>
<td>Positive</td>
<td>Bev + Erl (n=77) Erl (n=75)</td>
<td>69 (P=0.0157)</td>
<td>16.4 (P=0.0157)</td>
<td>47.0 (P=0.0157)</td>
</tr>
<tr>
<td>Rosell (33)</td>
<td>109</td>
<td>First-line</td>
<td>Positive (n=109)</td>
<td>Bev + Erl T790M+ (n=37) T790M− (n=72)</td>
<td>76.1 (P=0.03)</td>
<td>13.8 (P=0.048)</td>
<td>Not reached (P=0.996)</td>
</tr>
<tr>
<td>Otsuka (34)</td>
<td>24</td>
<td>EGFR-TKIs resistance</td>
<td>Positive (n=24)</td>
<td>Bev + Erl/Gef T790M+ T790M−</td>
<td>13 (P=0.53)</td>
<td>4.1 (P=0.048)</td>
<td>13.5 (P=0.996)</td>
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<td>0 (P=0.48)</td>
<td>3.3 (P=0.048)</td>
<td>15.1 (P=0.996)</td>
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<td></td>
<td></td>
<td></td>
<td></td>
<td>18 (P=0.996)</td>
<td>4.1 (P=0.048)</td>
<td>13.5 (P=0.996)</td>
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<tr>
<td>Saito (35)</td>
<td>226</td>
<td>First-line</td>
<td>Positive</td>
<td>Bev + Erl (n=112) Erl (n=114)</td>
<td>72.3 (P=0.0157)</td>
<td>16.9 (P=0.0157)</td>
<td>Not reached (P=0.0157)</td>
</tr>
</tbody>
</table>

BAC, bronchioloalveolar carcinoma; Erl, erlotinib; Bev, bevacizumab; Chemo, chemotherapy; Pan, panitumumab; Afa, afatinib; Gef, gefitinib; WT, wild-type; bevacizumab in these studies was administered at a dose of 15 mg/kg; erlotinib in these studies was administered at a dose of 150 mg/day; afatinib in these studies was administered at a dose of 40 or 30 mg/day.
erlotinib alone (16.0–16.9 vs. 9.7–13.3 months) (32,35). We also expect that the results of the ongoing BEVERLY clinical trials will further inform us as to the combined efficacy of bevacizumab and erlotinib (36). Interestingly, a subgroup analysis in the one-armed BELIEF study found that the de novo T790M-positive subgroup had a significantly prolonged median PFS compared to the T790M-negative subgroup (16.0 vs. 10.5 months) when patients were treated with first-line erlotinib combined to bevacizumab (33). The reason for this observed effect remains unclear.

Based on these aforementioned preclinical and clinical trials, it is clear that EGFR-TKIs combined with anti-angiogenic targeted drugs results in superior treatment outcomes compared to EGFR-TKIs alone. Our findings indicate that the anti-tumor efficacy on T790M positive lung adenocarcinoma of osimertinib combined with apatinib is significantly higher than that of osimertinib alone, in both in vitro and in vivo experiments. Apatinib is a potent small molecule VEGFR2 TKI used in second- and third-line treatment of patients with advanced NSCLC, that failed to respond to prior chemotherapy or EGFR-TKIs. Several clinical studies have shown that the ORR and disease control rate (DCR) of single drug apatinib, in the second-line of treatment or greater, in advanced NSCLC patients ranged from 8% to 26% and from 61.9% to 95.5%, respectively. The median PFS and OS ranged from 3 to 6.77 months and from 6 to 8.2 months, respectively (37-42).

Regarding apatinib combination therapy, several preclinical and clinical studies demonstrated an enhanced anti-tumor efficacy using a combination of apatinib and docetaxel for NSCLC, compared to drug monotherapy (43). Another preclinical study reported by Li et al. (44) demonstrated that gefitinib plus apatinib strengthened the anti-tumor effect of gefitinib or apatinib alone in H1975 cells, and that the combined administration delayed tumor growth in a xenograft model, and had a stronger effect on the inhibition of the activation of EGFR and VEGFR2 downstream pathway members, compared to drug monotherapy. These results are in accordance with the results of this study, which showed that apatinib monotherapy had a weak and negligible effect on anti-tumor activity in vitro, yet a strong synergetic anti-tumor effect in PC9 or H1975 cells when combined with osimertinib. These results suggest that osimertinib combined with apatinib treatment dually inhibits the EGFR and VEGFR signaling pathways. Lichtenberger et al. (45) have reported that there exists “crosstalk” between the EGFR and VEGF pathways, both of which have synergistic effects on tumor growth. VEGF is downregulated by EGFR inhibition, with one of the reasons for the development of EGFR-TKI resistance being increased expression of the VEGF protein. In the present study, we observed that combination therapy in vitro significantly increased the inhibition of EGFR downstream signaling pathway members PI3K-AKT and RAS-ERK. The in vivo synergetic inhibitory effect could be implemented by improving the tumor microenvironment, increasing the drug concentration of osimertinib in tumor tissues and enhancing the blockade of EGFR downstream signal transduction. We also found that a low concentration of osimertinib combined with apatinib had an efficacy similar to that of a high concentration of osimertinib. This is an important factor for drugs such as those with dose-limited toxicity, as combination therapy might result in a decreased and tolerable toxicity, thereby avoiding the severe adverse effects caused by simply increasing dosage.

Currently, there are a limited number of clinical studies that have investigated restoring EGFR-TKI sensitivity by adding an anti-angiogenic drug in combination with an EGFR-TKI. A study by Otsuka et al. (34) showed that EGFR-TKIs re-challenged with bevacizumab resulted in a modestly prolonged PFS and OS for patients with EGFR-mutant NSCLC that had acquired resistance to EGFR-TKI. The median PFS and OS for T790M-positive versus T790M-negative NSCLC was 3.3 vs. 4.1 months, and 15.1 vs. 13.5 months, respectively. Therefore, using EGFR-TKIs in combination with anti-angiogenic drugs to overcome or reverse EGFT-TKI resistance seems to be a promising strategy.

In the present study, before patients were enrolled, they had previously received multi-line chemotherapeutic regimens and acquired secondary resistance to both erlotinib and osimertinib, despite displaying initial responses to the two drugs. The patients had a PFS range of 10–39 months with erlotinib and 3–6 months with osimertinib, respectively (Table 1). According to our preclinical study, they received a combination therapy of osimertinib (80 mg/day) plus apatinib (250–500 mg/day). All patients responded to the combination therapy with a prolonged PFS ranging from 5 to 7 months, which is even longer than the median PFS obtained when these patients with the T790M mutation initially received osimertinib. Of note, these patients did not stop taking osimertinib after the failure of osimertinib; that is, there was no treatment interval or so-called osimertinib re-challenge. This suggests that the addition of apatinib to osimertinib may not only
enhance the synergistic anti-tumor effect observed in our preclinical study, but also partially restore a patient’s resistance to osimertinib. Of course, different osimertinib-resistance mechanisms might impact the efficacy of this combination therapy, therefore large clinical studies based on molecular detection are warranted. The most common grade 3 adverse events were fatigue, oral mucositis, and anorexia, which usually required symptomatic treatment to avoid interruption or discontinuity in the study.

In conclusion, apatinib can significantly increase the anti-tumor effect of osimertinib for EGFR T790M mutation-positive lung adenocarcinoma, by means of the synergic blockade of the activation of ERK and AKT signal transduction downstream of EGFR. The clinical application of osimertinib combined with apatinib as a treatment choice for osimertinib-resistant patients is promising. Our findings provide a strong basis for the design of clinical trials for this purpose.

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**Footnote**

**Conflicts of Interest:** All authors have completed the ICMJE uniform disclosure form (available at http://dx.doi.org/10.21037/tcr.2019.09.35). The authors have no conflicts of interest to declare.

**Ethical Statement:** The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. This study was conducted in accordance with the Declaration of Helsinki (as revised in 2013). This study was approved by the Ethics Committee of Shengjing Hospital of China Medical University (approval ID: 2015PS167K), and all of the patients have signed the informed consent forms.

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